



**University of
Zurich**^{UZH}

**Zurich Open Repository and
Archive**

University of Zurich
University Library
Strickhofstrasse 39
CH-8057 Zurich
www.zora.uzh.ch

Year: 2015

Hereditary diffuse gastric cancer: updated clinical guidelines with an emphasis on germline CDH1 mutation carriers

van der Post, Rachel S ; Vogelaar, Ingrid P ; Carneiro, Fátima ; et al

Abstract: Germline CDH1 mutations confer a high lifetime risk of developing diffuse gastric (DGC) and lobular breast cancer (LBC). A multidisciplinary workshop was organised to discuss genetic testing, surgery, surveillance strategies, pathology reporting and the patient's perspective on multiple aspects, including diet post gastrectomy. The updated guidelines include revised CDH1 testing criteria (taking into account first-degree and second-degree relatives): (1) families with two or more patients with gastric cancer at any age, one confirmed DGC; (2) individuals with DGC before the age of 40 and (3) families with diagnoses of both DGC and LBC (one diagnosis before the age of 50). Additionally, CDH1 testing could be considered in patients with bilateral or familial LBC before the age of 50, patients with DGC and cleft lip/palate, and those with precursor lesions for signet ring cell carcinoma. Given the high mortality associated with invasive disease, prophylactic total gastrectomy at a centre of expertise is advised for individuals with pathogenic CDH1 mutations. Breast cancer surveillance with annual breast MRI starting at age 30 for women with a CDH1 mutation is recommended. Standardised endoscopic surveillance in experienced centres is recommended for those opting not to have gastrectomy at the current time, those with CDH1 variants of uncertain significance and those that fulfil hereditary DGC criteria without germline CDH1 mutations. Expert histopathological confirmation of (early) signet ring cell carcinoma is recommended. The impact of gastrectomy and mastectomy should not be underestimated; these can have severe consequences on a psychological, physiological and metabolic level. Nutritional problems should be carefully monitored.

DOI: <https://doi.org/10.1136/jmedgenet-2015-103094>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-119259>

Journal Article

Published Version



The following work is licensed under a Creative Commons: Attribution-NonCommercial 4.0 International (CC BY-NC 4.0) License.

Originally published at:

van der Post, Rachel S; Vogelaar, Ingrid P; Carneiro, Fátima; et al (2015). Hereditary diffuse gastric cancer: updated clinical guidelines with an emphasis on germline CDH1 mutation carriers. *Journal of Medical Genetics*, 52(6):361-374.

DOI: <https://doi.org/10.1136/jmedgenet-2015-103094>



OPEN ACCESS

Hereditary diffuse gastric cancer: updated clinical guidelines with an emphasis on germline *CDH1* mutation carriers

Rachel S van der Post,¹ Ingrid P Vogelaar,² Fátima Carneiro,^{3,4,5,6} Parry Guilford,⁷ David Huntsman,⁸ Nicoline Hoogerbrugge,² Carlos Caldas,⁹ Karen E Chelcun Schreiber,¹⁰ Richard H Hardwick,¹¹ Margreet G E M Ausems,¹² Linda Bardram,¹³ Patrick R Benusiglio,¹⁴ Tanya M Bisseling,¹⁵ Vanessa Blair,¹⁶ Eveline Bleiker,¹⁷ Alex Boussioutas,^{18,19} Annemieke Cats,²⁰ Daniel Coit,²¹ Lynn DeGregorio,²² Joana Figueiredo,^{3,4} James M Ford,²³ Esther Heijkoop,²⁴ Rosella Hermens,²⁵ Bostjan Humar,²⁶ Pardeep Kaurah,²⁷ Gisella Keller,²⁸ Jennifer Lai,¹⁰ Marjolijn J L Ligtenberg,^{1,2} Maria O'Donovan,²⁹ Carla Oliveira,^{3,4,5} Hugo Pinheiro,^{3,4} Krish Ragunath,³⁰ Esther Rasenberg,³¹ Susan Richardson,³² Franco Roviello,³³ Hans Schackert,³⁴ Raquel Seruca,^{3,4,5} Amy Taylor,³⁵ Anouk ter Huurne,³⁶ Marc Tischkowitz,³⁷ Sheena Tjon A Joe,²³ Benjamin van Dijk,³⁶ Nicole C T van Grieken,³⁸ Richard van Hillegersberg,³⁹ Johanna W van Sandick,⁴⁰ Rianne Vehof,⁴¹ J Han van Krieken,¹ Rebecca C Fitzgerald^{42,43,44}

► Additional material is published online only. To view please visit the journal online (<http://dx.doi.org/10.1136/jmedgenet-2015-103094>).

For numbered affiliations see end of article.

Correspondence to

Professor Rebecca C Fitzgerald, MRC Cancer Unit, Hutchison/MRC Research Centre, University of Cambridge, Hills Road, Cambridge CB2 0XZ, UK; rcf29@MRC-CU.cam.ac.uk

RSvdP and IPV are joint first authors.

Received 26 February 2015
Accepted 18 March 2015



Open Access
Scan to access more
free content



CrossMark

To cite: van der Post RS, Vogelaar IP, Carneiro F, et al. *J Med Genet* 2015;**52**:361–374.

ABSTRACT

Germline *CDH1* mutations confer a high lifetime risk of developing diffuse gastric (DGC) and lobular breast cancer (LBC). A multidisciplinary workshop was organised to discuss genetic testing, surgery, surveillance strategies, pathology reporting and the patient's perspective on multiple aspects, including diet post gastrectomy. The updated guidelines include revised *CDH1* testing criteria (taking into account first-degree and second-degree relatives): (1) families with two or more patients with gastric cancer at any age, one confirmed DGC; (2) individuals with DGC before the age of 40 and (3) families with diagnoses of both DGC and LBC (one diagnosis before the age of 50). Additionally, *CDH1* testing could be considered in patients with bilateral or familial LBC before the age of 50, patients with DGC and cleft lip/palate, and those with precursor lesions for signet ring cell carcinoma. Given the high mortality associated with invasive disease, prophylactic total gastrectomy at a centre of expertise is advised for individuals with pathogenic *CDH1* mutations. Breast cancer surveillance with annual breast MRI starting at age 30 for women with a *CDH1* mutation is recommended. Standardised endoscopic surveillance in experienced centres is recommended for those opting not to have gastrectomy at the current time, those with *CDH1* variants of uncertain significance and those that fulfil hereditary DGC criteria without germline *CDH1* mutations. Expert histopathological confirmation of (early) signet ring cell carcinoma is recommended. The impact of gastrectomy and mastectomy should not be underestimated; these can have severe consequences on a psychological, physiological and metabolic level. Nutritional problems should be carefully monitored.

INTRODUCTION

Worldwide, gastric cancer (GC) is the fifth leading cause of cancer and the third most common cause

of death from cancer, with an estimated number of 723 000 deaths annually.¹ The vast majority of GCs are sporadic, but it has now been established that 1–3% of GCs arise as a result of inherited cancer predisposition syndromes. These syndromes include Li-Fraumeni syndrome,^{2–4} Lynch syndrome,^{5–7} Peutz-Jeghers syndrome,^{8–10} hereditary breast and ovarian cancer,^{11 12} *MUTYH*-associated adenomatous polyposis (MAP),¹³ familial adenomatous polyposis,^{14–16} juvenile polyposis syndrome^{17 18} and *PTEN* hamartoma tumour syndrome (Cowden syndrome).¹⁹ The lifetime risk of GC in these syndromes varies substantially between populations studied, but is generally low.

Over 15 years ago, linkage analysis implicated germline mutations in the *CDH1* gene, encoding the tumour-suppressor protein E-cadherin, as the genetic cause of hereditary diffuse GC (HDGC).²⁰ Heterozygous germline *CDH1* mutations increase lifetime risk of developing diffuse GC (DGC) and lobular breast cancer (LBC). Criteria have been set to select families eligible for screening of germline *CDH1* mutations, and they were updated in 2010.^{21 22} Not all families fulfilling these criteria have mutations in *CDH1*, indicating that other genes may also be involved in DGC predisposition. Germline mutations in *CTNNA1* were described in three families that presented with DGC, one of them fulfilled the 2010 HDGC criteria.^{23 24}

Increasing awareness of HDGC and the rapid advances in genetic diagnostic tools, endoscopic modalities and the increasing use of laparoscopic surgery led a group of clinical geneticists, gastroenterologists, surgeons, oncologists, pathologists, molecular biologists, dieticians and patients' representatives from nine different countries to convene a workshop in order to update the management guidelines for this condition set in 2010 and to

Review

propose directions for future research. The workshop discussions were focused on five major topics: (1) genetic counselling and mutation analysis; (2) endoscopic surveillance and screening of cancer; (3) risk-reduction surgery of the stomach and breasts; (4) pathological specimen processing and diagnosis; and (5) patients' and dieticians' perspective.

GENETIC COUNSELLING AND MUTATION ANALYSIS

Genetic evaluation of patients with HDGC

Genetic counselling is an essential component of the evaluation and management of HDGC. The counselling process should include a formal genetics evaluation by a cancer genetics professional with expertise in the field. The evaluation should include a detailed three-generation family pedigree, histopathological confirmation of DGC diagnoses and/or precursor lesions (*in situ* or pagetoid spreading of signet ring cells) and a discussion on lifetime risks of developing DGC and LBC. Informed consent for genetic testing is required. In the management of individuals with a *CDH1* mutation, a full multidisciplinary team (MDT) should be involved comprising those with relevant expertise in gastric surgery, gastroenterology, breast oncology, pathology, psychosocial support and nutrition. Genetic testing can be offered from the age of consent (and therefore will vary between countries, but will generally be around 16–18 years). Testing of younger unaffected family members can be considered on a case-by-case basis. Rare cases of clinically significant DGC have been reported in affected families before the age of 18,²⁵ but the overall risk of this disease before the age of 20 is low.^{26 27}

Cancer risks in *CDH1* mutation carriers

In a recent study, penetrance data for *CDH1* mutation carriers has been updated based on affected individuals, who presented clinically with DGC or LBC, from 75 families with pathogenic *CDH1* mutations. Families with *CDH1* missense mutations and families for which no carrier test information was available were excluded from this analysis. The cumulative risk of DGC for *CDH1* mutation carriers by age 80 years is reported to be 70% for men (95% CI 59% to 80%) and 56% for women (95% CI 44% to 69%). Furthermore, the cumulative risk of LBC for women with a *CDH1* mutation is estimated to be 42% (95% CI 23% to 68%) by 80 years. There is currently no evidence that the risk of other cancer types in individuals with a *CDH1* mutation is significantly increased.²⁴

Implications of counselling

Clinical geneticists (or other members of the MDT) should inform the counsellee about gastric surgery and options of surveillance. Proven pathogenic *CDH1* germline mutation carriers should be advised to undergo prophylactic gastrectomy (for more details, see the 'Surgery' section). Some patients may want to delay prophylactic gastrectomy for personal and/or work-related reasons. In these individuals, endoscopic surveillance, described in detail under the 'Endoscopic surveillance' section, should be considered in the interim. The management of patients and families with clinical features suggestive of HDGC, but without a germline *CDH1* mutation, is not straightforward. We advise intensive endoscopic surveillance in an expert centre for first-degree relatives of patients meeting criteria mentioned in figure 1. This is also the case for patients and families who

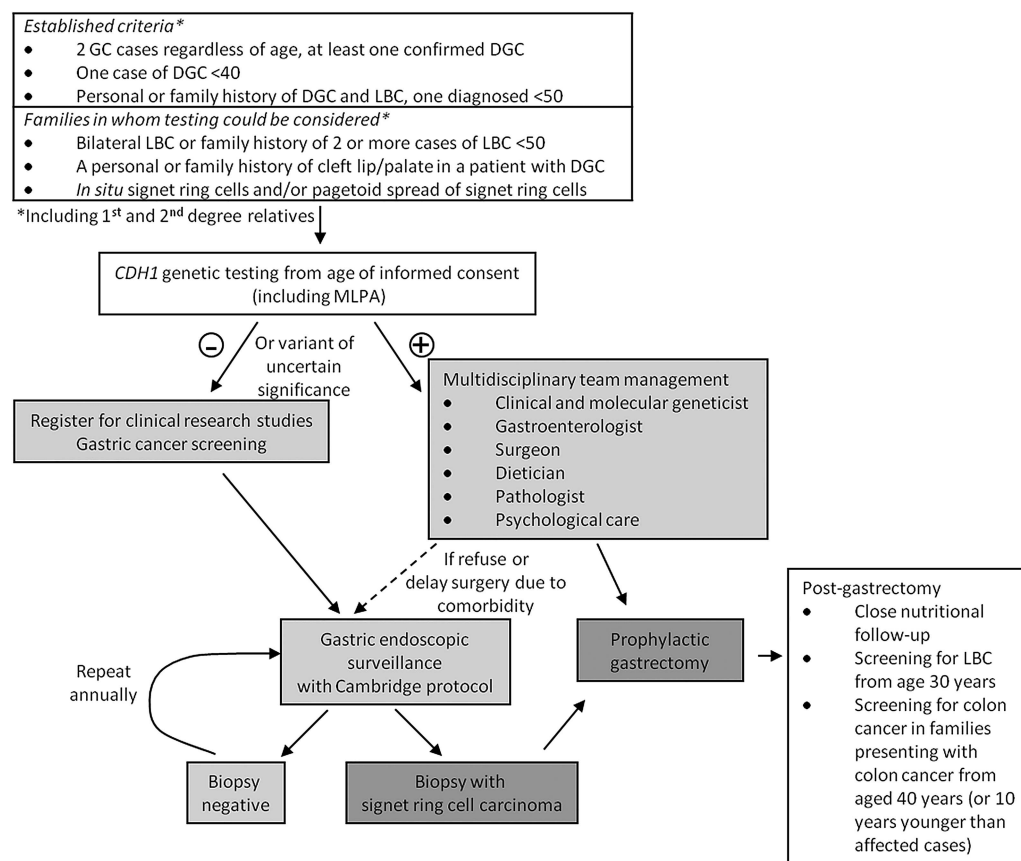


Figure 1. Algorithm for management starting from clinical hereditary diffuse gastric cancer (HDGC) testing criteria, genetic testing, role of endoscopy and gastrectomy. GC, gastric cancer; DGC, diffuse gastric cancer; LBC, lobular breast cancer; MLPA, multiplex-ligation probe amplification.

carry a *CDH1* variant with unproven deleterious effect. Recommendations on breast cancer surveillance and therapy in *CDH1* mutation carriers can be found below.

Criteria for germline *CDH1* mutation testing

The last guideline in 2010 broadened the clinical criteria to select patients eligible for *CDH1* mutation analysis. The detection rate of *CDH1* mutations before 2010, using the guidelines established in 1999,²¹ was reported to be 25–50%.^{27–28} Using the new criteria, the *CDH1* detection rate in countries with a low incidence of GC has decreased to 10–18% (RS van der Post *et al*, under revision, 2015).^{24–29} This decrease may also be explained partly by the enrichment of large and highly suspected families in studies published before 2010 (ie, ascertainment bias). The detection rate is considerably higher in the New Zealand Maori population and is likely higher in the Newfoundland population, where a founder mutation has been described.²⁷ It should be noted that these criteria are designed for countries with a low incidence of sporadic GC. Few small series and case reports of individuals with *CDH1* mutations have been described in countries with a high incidence of sporadic GC; no large cohorts were examined systematically. In small series of South Korea, Japan and Portugal, mutation detection rates of pathogenic germline *CDH1* mutations in HDGC and early-onset GC range between 8% and 15%.^{2–30–33}

During the workshop, it was decided to merge the first ('Two or more GC cases in a family, one DGC <50') and second ('Three or more DGC cases regardless of age') testing criteria from the previous guideline²² into a new criterion: 'Two or more GC cases regardless of age, at least one confirmed DGC', in first-degree and second-degree relatives. The two other testing criteria have not been changed. The revised criteria are depicted in the upper panel of the box in [figure 1](#). The revised criterion now covers families for whom detailed pathology is incomplete. However, as in the earlier criteria, confirmed intestinal-type GC cases are not part of HDGC and in these families no *CDH1* mutation analysis should be performed. To properly assess whether a family meets the HDGC criteria, pathology reports and preferably review of gastric specimens by a pathologist dedicated to GC are essential.

There are other families in which genetic testing may also be considered ([figure 1](#), lower panel). The presence of bilateral LBC at young age (under the age of 50 years) or the presence of multiple close relatives with LBC (at least two under the age of 50 years) may justify testing for *CDH1* mutations.^{29–34} Testing should also be considered in individuals with a personal or family history of cleft lip/cleft palate and DGC.^{35–36} Furthermore, in cases where expert pathologists detect *in situ* signet ring cells and/or pagetoid spread of signet ring cells in the stomach, genetic testing should be considered since this is rarely (if ever) seen in sporadic cases. Genetic testing should, whenever possible, be initiated in an affected proband. If the affected proband is deceased, frozen-fixed or formalin-fixed, paraffin-embedded tissue (preferably normal, non-malignant tissue) still may be available for *CDH1* germline mutation testing. In case tissue of an affected individual is not available or of insufficient quality to perform the test, testing of unaffected relatives is acceptable. This should be performed preferably in at least three first-degree relatives simultaneously, thus increasing the likelihood of detecting a *CDH1* mutation. However, it has to be made clear to the counselee that unaffected individuals have only a 50% chance of having inherited a mutated *CDH1* allele. The testing criteria still have to be met by the counselee's family. If no mutation is found in these cases, endoscopic

screening should be discussed with an expert gastroenterologist on a case-by-case basis. Similarly, in small families with DGC or in families where family history is unavailable, *CDH1* mutation screening should also be considered on a case-by-case basis.

Genetic testing: lab perspective

Genetic testing on blood or tissue for germline mutations should be performed in certified molecular diagnostic laboratories, for example, Clinical Laboratory Improvement Amendments approved, ISO 15189 accredited or equivalent. *CDH1* analysis should include mutation analysis of the entire open-reading frame, including intron–exon boundaries, as well as copy number analysis of individual exons to detect intragenic exon deletions or duplications. At present, analyses are mostly performed by a combination of Sanger sequencing and Multiplex Ligation-dependent Probe Amplification. These techniques can be replaced by next-generation sequencing if the approach used fulfils the requirement of similar sensitivity.

To date, over 155 different germline *CDH1* mutations have been described, the majority pathogenic mutations and a number of variants of uncertain significance (RS van der Post *et al*, under revision, 2015).²⁴ The majority of the pathogenic mutations are truncating and thus do not lead to a functional protein. Large exonic deletions are relatively rare, with a frequency of about 5%.^{37–38} *CDH1* is a tumour suppressor gene and therefore a somatic second hit is required for initiation of tumour formation. The trigger and molecular mechanism by which the second allele of *CDH1* is inactivated appears to be diverse and includes methylation, somatic mutation and loss of heterozygosity.^{39–42}

The identification of germline *CDH1* missense variants requires additional studies to assess their putative pathogenicity. A multidisciplinary approach combining familial and population data, *in silico* analysis and *in vitro* analysis is currently used to classify the variants as neutral or pathogenic.^{43–47} Genetic parameters such as mutation frequency in healthy control populations, co-segregation of the mutation within the pedigree and recurrence of the mutation in independent families should be considered as a first approach.^{43–45} However, the low number of individuals affected by the disease, the small size of the pedigrees and the absence of mutation hotspots (which prevents the establishment of any correlation between the mutation site and its functional consequence) are limitations of this approach. *In silico* experiments predict the degree of conservation of a specific nucleotide within species, the effect of missense variants on splicing and also the putative impact of the variants on protein structure.^{45–47} Still, *in silico* results should be carefully interpreted because the degree of conservation among species of each amino acid position is considered separately and does not take into account possible compensatory effects of neighbouring amino acids.^{45–47} Functional *in vitro* assays should be performed in order to evaluate the impact of *CDH1* missense alterations in protein structure, trafficking, signalling and, consequently, in E-cadherin function.^{44–46} When compared with the cells expressing wild-type E-cadherin, pathogenic missense mutants impair the correct binding of key adhesion-complex regulators and likely compromise normal E-cadherin localisation and stability at the plasma membrane.^{46–48} As a consequence, disruption of cell–cell adhesion and increased invasive behaviour may be observed in the presence of pathogenic variants.^{44–46} To date, 49 germline *CDH1* missense variants were reported for functional evaluation to Institute of Molecular Pathology and Immunology of the University of Porto (Portugal, reference laboratory for functional characterisation of *CDH1* missense

Review

variants for the International Gastric Cancer Linkage Consortium), the majority being classified as deleterious and thus possibly pathogenic (R Seruca, personal communication, 2014).

Panel sequencing

Penetrance estimates for *CDH1* mutation carriers have been derived from the study of highly ascertained HDGC families and it is likely that the penetrance for mutations detected in non-HDGC families will be lower. With the introduction of next-generation sequencing-based gene panels, both in research and diagnostic settings, *CDH1* alterations may be found in patients without a personal or family history of GC.^{49 50} In our opinion, one should be very cautious in the interpretation of coding variants identified in non-HDGC families, especially if the alterations do not lead to a premature stop codon.

CDH1 mutation database

Currently, there is no international database containing all germline *CDH1* mutations and variants identified to date. A database has been designed and is currently under construction with the collaboration of the LOVD team. The variant database is available at <http://www.LOVD.nl/CDH1>. This database can be consulted to assess whether a given *CDH1* mutation has been found by others and whether it has been considered deleterious and likely disease-causative or not based on population data, segregation analysis, *in silico* analysis and *in vitro* functional analysis, and/or recurrence in several individuals/families. We advise researchers and clinicians to submit unpublished mutations and variants to the database (contact C Oliveira, carlaol@ipatimup.pt), together with the requested information on families/patients and mutations. The publication/submitter of every mutation will always be referred to in the database.

Psychosocial effects of counselling

Even though it is well recognised that many individuals will benefit from genetic counselling and testing for hereditary cancer in general, there have also been reports that it may induce a number of psychosocial problems. In a review on individuals requesting genetic counselling and testing for hereditary cancer syndromes, six dominant problem areas were identified: (1) coping with cancer risk; (2) practical problems (such as obtaining life insurance/loans and employment when found to be a mutation carrier); (3) family-related problems (eg, communication problems with family members, feeling responsible for family members); (4) children-related problems (eg, concerns for children having increased risk, fear of leaving young children); (5) living with cancer (eg, fear of developing cancer, pain about the loss of family members) and (6) emotions (eg, anxiety, anger, feelings of loss, but also relief and reassurance).⁵¹ These topics, when applicable, should be addressed during the counselling sessions.

Pregnancy and assisted reproduction

Although scientific data are lacking concerning timing of prophylactic gastrectomy and family planning, it is entirely possible for women to give birth to a healthy child after gastrectomy.⁵² Nutritional advice and follow-up with a dietician within this context is essential.

Individuals from hereditary cancer families are frequently concerned about the transmission of their predisposition of cancer to their children.^{53 54} Healthcare professionals, including geneticists and psychosocial workers, will be increasingly involved in discussions and decisional counselling regarding

reproductive options in families with a known cancer predisposing mutation such as *CDH1*. In the past decades, genetic testing for hereditary cancers before birth has become available through prenatal diagnosis (PND) and preimplantation genetic diagnosis (PGD).⁵⁵ We recommend that carriers of a *CDH1* mutation with a desire to have children should be informed about all reproductive options, including PND and PGD.

Future research: other genes involved in HDGC predisposition

Currently, three families that meet the new criteria have been described to carry germline *CTNNA1* mutations.^{23 24} Even though these families show a clinical picture similar to that of *CDH1* mutation-positive families,²³ insufficient data are available to make a statement on disease penetrance. Given the functional connection between the two genes, they may represent a genocopy. Mutation carriers could be given the option of prophylactic total gastrectomy (PTG) and other cancer prevention measures recommended for HDGC families, but with the precaution that such advice is being given based on very limited data.

Other families have recently been described with *BRCA2* and *PALB2* mutations;²⁴ however, we recommend that these families are managed no differently than other families with such mutations according to national guidelines. It is likely that other HDGC-associated genes will be discovered through whole exome, genome or other unbiased next-generation-sequencing empowered methodologies. Indeed, using a combination of this approach and linkage analysis, mutations in *MAP3K6* have recently been described.⁵⁶ More needs to be understood about families with *MAP3K6* mutations before they could be used to stratify risk in families. Until such data are available, a cautious approach in which all first-degree relatives of mutation carriers are followed is recommended. Without multiple mutation-positive families for newly identified genes, it will be extremely difficult to ascribe pathogenicity to such mutations and to develop management guidelines.

SCREENING AND SURVEILLANCE

Gastric endoscopic screening and surveillance

To clarify the terminology, we consider that individuals having endoscopy who do not know their mutation status or those who do not have a proven pathogenic *CDH1* mutation undergo screening whereas mutation-positive individuals undergo surveillance. The consensus reached at the workshop was that individuals who tested positive for a pathogenic germline *CDH1* mutation should be advised to consider prophylactic gastrectomy, regardless of endoscopic findings. However, the timing of surgery may vary according to the preferences and age, as well as the physical and psychological fitness of the individual. In patients proceeding for gastrectomy, a baseline endoscopy should be performed prior to surgery to look for macroscopic tumour as this may alter the treatment plan. This endoscopy is also performed to ensure that there is no other coincidental pathology, such as Barrett's oesophagus, which may alter the extent of the resection. When the stomach is macroscopically normal, the information on microscopic disease *foci* is useful to compare with findings in the surgical resection specimen and hence to increase knowledge on the likelihood of endoscopic detection of microscopic lesions.

For individuals with a *CDH1* mutation in whom gastrectomy is not currently being pursued (eg, through patient choice or existence of physical or psychological comorbidity), regular endoscopy should be offered. In patients declining surgery, surveillance can have the advantage of helping individuals to come

to a decision about the need for gastrectomy when microscopic *foci* of signet ring cells are detected. However, patients should be aware that delaying surgery can be a hazardous decision.⁵⁷

The management of individuals with a *CDH1* variant of uncertain significance and those in whom no mutation can be identified in the family is not straightforward. We would recommend that intensive endoscopic surveillance in an expert centre should be offered to these families who fulfil the HDGC criteria. Endoscopic screening has a valuable role in guiding clinical decision making and in one case series lesions were detected in 2/7 *CDH1* mutation-negative individuals (1/5 families).⁵⁷ Specifically, any malignant lesions detected endoscopically would prompt a referral for gastrectomy. However, all patients undergoing endoscopy for HDGC should be informed that, given the very focal and often endoscopically invisible nature of these lesions, it is quite possible that lesions will not be detected by random biopsies.

HDGC endoscopy protocol

Endoscopy should be performed in centres with an experienced MDT. However, it is appreciated that sometimes this is not practical for individuals who have to travel long distances. In this case, a local endoscopist in consultation with an expert centre on the endoscopy protocol and review of histology may be a helpful alternative.

As noted above, the optimal frequency of endoscopy is not known. Based on current experience, it is recommended that individuals should be offered annual endoscopy. The bleeding risk may be slightly higher than for other indications since more biopsies are taken. Therefore, it is recommended that the local high-risk endoscopy protocol is followed such that, if possible, anticoagulants (eg, warfarin and clopidogrel) are stopped prior to the procedure. The endoscopy should be performed using a white light high definition endoscope in a dedicated session of at least 30 min to allow for careful inspection of the mucosa on repeated inflation and deflation and for collection of biopsies. The mucosa should be thoroughly washed before examination with a combination of mucolytics (N-acetylcysteine) and anti-foaming agent (such as simethicone) mixed with sterile water. This washing is ideally done via a pump operated by a foot pedal. The macroscopic appearances of the gastric mucosa and any focal visible lesions should be recorded using still images or video for future reference and specifically sampled for histology prior to the collection of random biopsies.

Prior to examination for small *foci*, the stomach should be adequately inflated and deflated to check distensibility. Poor distensibility should raise alarm for a submucosal infiltrative process like *limitis plastica*. When this is the case, biopsies should be taken and further imaging such as a high-resolution multidetector CT scan combined with endoscopic ultrasonography is suggested to visualise the gastric wall layers. No objective measures of distensibility are currently available, and this is an area that may warrant future research.

Although an association between *Helicobacter pylori* infection and HDGC has not been proven, it is important to test for *H. pylori* to document the prevalence of infection. Since *H. pylori* is a WHO class 1 carcinogen, it is agreed that when individuals are infected it should be eradicated, especially in those opting for surveillance. A rapid urease test is the preferred test at baseline, and additionally, it is recommended to take random biopsies from the antrum and the corpus due to patchy colonisation, especially in the presence of acid suppression.

Due to the tiny *foci* of signet ring cells, which can only be recognised by microscopy, multiple biopsies are required to

maximise the likelihood of diagnosing them.³⁹ The anatomical gastric localisation in which *foci* are identified varies between studies; reasons for this remain to be clarified but may include environmental factors or differences in the molecular pathogenesis.^{39 57–65} Therefore, it is recommended that any endoscopically visible lesions are biopsied including pale areas. Additionally, random sampling should be performed comprising five biopsies taken from each of the following anatomical zones: pre-pyloric area, antrum, transitional zone, body, fundus and cardia. A minimum of 30 biopsies is recommended as described in the Cambridge protocol (see online supplementary protocol 1).²² Even though this will still lead to sampling bias due to the large gastric surface area, taking more biopsies is not feasible in practice.⁶⁵ The biopsies may be taken using a standard forceps, ideally with a spike as this will seize the lamina propria in which signet ring cell *foci* are present. In the case of a well-defined visible lesion, an endoscopic mucosal resection can be helpful to achieve a more reliable histopathological specimen to document the degree of invasion. However, this should be done for diagnostic rather than therapeutic purposes in view of the multifocal nature of the lesions.

Special mention should be given to pale areas since these are more likely to harbour microscopic *foci* of abnormal cells, although they lack specificity leading to false positives (figure 2).⁶⁶ Recent data also suggest that these areas are visible on careful examination by white light, but narrow band imaging may make them easier to visualise (A Cats, personal communication, 2014). As noted in the previous guidelines, chromoendoscopy with Congo-red and methylene blue is no longer recommended due to concerns over toxicity.⁶⁶ Virtual chromoendoscopy using autofluorescence and trimodal imaging does not seem to confer much additional benefit over white light.⁵⁷ In order to maximise the yield from endoscopy, specialist histopathology reporting is essential and the guidelines outlined in the pathology section below should be followed.

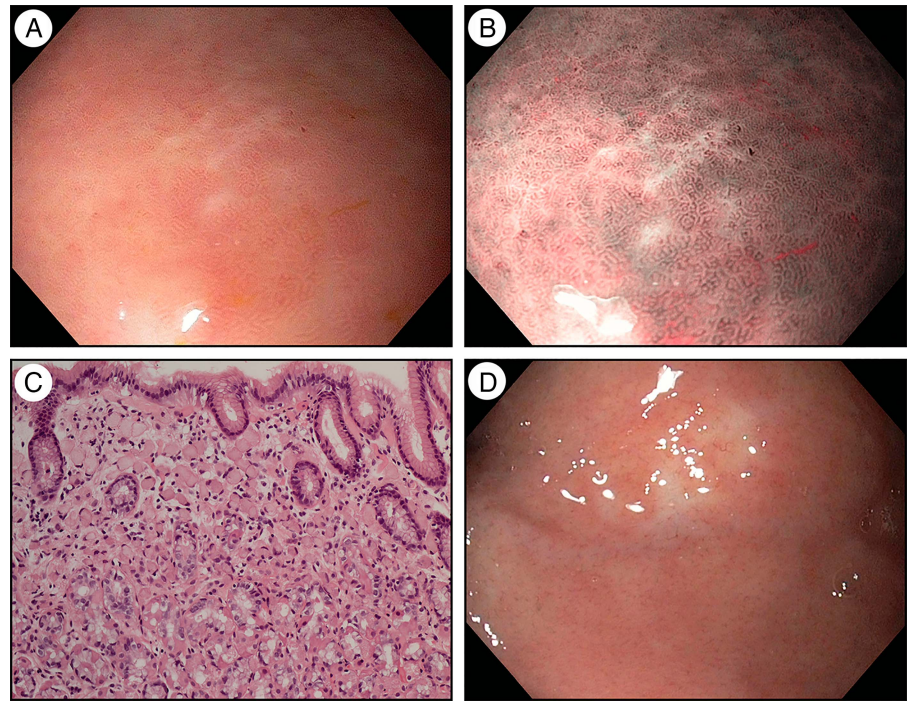
Endoscopic surveillance of colorectal cancer

Although there are case reports of colorectal and appendiceal signet ring cell carcinomas (SRCCs) in *CDH1* mutation carriers,^{26 67–70} there is currently no evidence to suggest that the risk of colorectal cancer in *CDH1* mutation carriers is significantly elevated and there are insufficient data to give recommendations on colorectal cancer screening. In *CDH1* mutation families in which colon cancer is reported in mutation carriers, information should be collected concerning the age at diagnosis, whether the affected member(s) and first-degree or second-degree relatives are mutation carriers and whether the histopathology showed a mucinous component and/or signet ring cells. For such families, enhanced colonoscopy screening should be considered at age 40 or 10 years younger than the youngest diagnosis of colon cancer, whichever is younger, and repeated at intervals of 3–5 years. In the absence of a family history, the national guidelines for colon cancer screening should be followed. It is imperative that data on colonoscopic screening in these individuals are collected so that these guidelines can be based more on evidence than on specialist opinion in the future.

Breast cancer surveillance

Knowledge about breast cancer risk in HDGC has slowly advanced since first reported in 2000,²⁶ yet evidence is not sufficient such that recommendations can be made of comparable strength as in *BRCA1/2*. Genotype–phenotype correlations may eventually show some HDGC families do not have an increased LBC risk, but at present it should be assumed all women with a

Figure 2 Pale areas in gastric mucosa of a patient with a germline *CDH1* mutation harbouring signet ring cell focus during white light endoscopy (A) and narrow band imaging (B). H&E stain of biopsy specimen with signet ring cells (C). Scar area after biopsy during previous endoscopy (D).



CDH1 mutation are at risk. When informing women with *CDH1* mutation about the role of breast surveillance versus prophylactic mastectomy to manage their risk, the consultation covers similar territory as in *BRCA1/2*. There are, however, some important differences that must be carefully explained.

Invasive LBC, the type that typically occurs in HDGC, makes up 5–15% of sporadic breast cancer. Invasive ductal cancer occurs in 85–95% of sporadic breast cancer and in *BRCA1/2* cases.⁷¹ In contrast to ductal breast cancer, E-cadherin-deficient LBC invades in sheets or cords of cells, typically in single file, and does not form a well-defined mass. Studies comparing different radiological modalities in sporadic LBC show sensitivity on mammography ranges from 34% to 92%.⁷¹ Bilateral breast MRI needs to be part of the protocol in *CDH1* mutation carriers. Given the fact that mammography has a low sensitivity for LBC, synchronous mammography at the time of MRI may add little. We therefore recommend annual breast MRI (which can be combined with mammography) starting at age 30 in women with a *CDH1* mutation. An oncologist or breast surgeon should guide breast surveillance. Annual clinical breast examination and breast cancer awareness from the patient and her physicians are essential.

Future research on gastric and breast surveillance

In light of emerging endoscopic technologies, such as narrow band imaging, blue laser imaging, I scan, autofluorescence imaging, IHB-enhancement and confocal endomicroscopy,⁷² research is required to further study the optimal methods for endoscopic monitoring of individuals at risk for DGC. Given the rarity of this condition, these need to be multicentre studies with strictly defined protocols. In addition, it is likely that the multiple biopsy protocol leads to scarring that can masquerade as pale areas (figure 2D). It would therefore be very valuable to compare the yield of a targeted biopsy approach only with the current Cambridge protocol, which also includes multiple random biopsies. Such studies could also inform on the interobserver variation in the identification of pale areas and help define features indicative of a signet ring cell focus. An

endoscopic atlas created by endoscopists performing regular endoscopies on *CDH1* mutation carriers would be very useful to help standardise protocols and improve lesion recognition.

There are no studies specifically addressing screening for LBC. Trials on breast screening in the general population and MRI screening in high-risk groups or *BRCA1/2* are informative but do not directly extrapolate to the screening scenario in HDGC. The outcomes of the above-stated recommendations of breast surveillance in HDGC women should be further prospectively investigated.

GASTRECTOMY AND MASTECTOMY

Prophylactic gastrectomy: indications for and timing of surgery

Prophylactic gastrectomy should be strongly advised in carriers of a proven pathogenic germline *CDH1* mutation. Some argue that the term ‘prophylactic gastrectomy’ is inaccurate and favour the term ‘risk reduction gastrectomy’ because most mutation carriers already have microscopic SRCCs at the time of their surgery. Total gastrectomy for these patients, however, completely eliminates their risk of GC and is truly prophylactic in terms of preventing their death from invasive GC.

The optimal timing of prophylactic gastrectomy is unknown and is usually highly individualised. Since this surgery has major impact on the quality of life, the decision to undergo prophylactic gastrectomy should be well informed, balanced, prepared and timed. Decisional counselling, outweighing the pros and the cons of the intervention is essential. The current consensus is that the procedure should be discussed and offered to pathogenic germline *CDH1* mutation carriers in early adulthood, generally between ages 20 and 30.⁵⁸ Based upon the physical fitness of the mutation carrier and of surgery-related complications, prophylactic gastrectomy at an age >75 should be carefully considered. Family phenotype, especially age of onset of clinical cancer in probands, should be taken into account. There is likely to be a dormant period in which the signet ring cell adenocarcinoma does not spread or progress since they have a low proliferative index and the age of prophylactic gastrectomy

is generally lower than that of overt cancer.³⁹ This may explain why so many individuals are found to have T-1 N-0 stage tumours after prophylactic gastrectomy.⁷³ Patients who develop symptomatic, widely invasive DGC have a poor prognosis with as few as 10% having potentially curable disease.⁷⁴ Even if potentially curable, 5-year survival rate still does not exceed 30%.⁷⁵ As our understanding of the natural history of mucosal SRCC improves, it may be possible to safely postpone prophylactic gastrectomy in some patients, but until such time it is safer to recommend surgery early in adult life. This has implications for the long-term follow-up of patients with prophylactic gastrectomy and reinforces the need for MDTs to care for these patients for the rest of their lives, similar to patients who have undergone bariatric malabsorptive surgery for obesity.⁷⁶

Operation details

The requisite operation is a total gastrectomy with Roux-en-Y reconstruction, ensuring that the jejunojejunal anastomosis is at least 50 cm distal to the oesophagogastric anastomosis, to reduce the risk of biliary reflux. The proximal resection line must be across the distal oesophagus containing squamous epithelium to ensure that no gastric cardiac mucosa is left behind. This can be confirmed by frozen section or examination of the opened resection specimen in operating room, and can be guided by the use of on-table endoscopy to mark the squamocolumnar junction during the surgery.

The optimal extent of lymph node dissection (LND) in prophylactic gastrectomy is controversial.⁷⁷ Lymph node metastases are not reported in asymptomatic *CDH1* mutation carriers with negative preoperative surveillance biopsies or small *foci* of pT1a intramucosal carcinoma. Among patients with early-stage intestinal adenocarcinoma of the stomach, the frequency of lymph node metastasis in patients with early intramucosal (pT1a) tumours is 2–5%,^{78–81} and up to 6% in the undifferentiated or diffuse types.^{78–80} In pT1b tumours, with invasion of the submucosal layer, lymph node metastases are found in 17–28%, increasing with the depth of submucosal invasion.^{80–81} The majority of patients undergoing prophylactic gastrectomy for HDGC will have at least T1a cancers. Because a preoperative gastroduodenoscopy cannot exclude the presence of T1b lesions with their higher risk of metastases during the operation, a D1 LND (with the inclusion of lymph node stations 1–7) is reasonable.

The formation of a jejunal pouch may improve eating for the first year after surgery, but, as yet, prospective trials comparing pouch to straight Roux-en-Y reconstruction have not convincingly demonstrated significant long-term benefits to justify the routine application of this more complex reconstructive procedure.⁸² Surgeons should therefore use the reconstruction they are most familiar with. This also applies to whether the preferred surgical approach is open or laparoscopic.^{77–83} There are potential advantages of laparoscopic gastrectomy with reduced wound pain and faster overall return to full activities, but conclusive evidence for the superiority of this approach is still lacking. Any surgeon proposing to do a laparoscopic PTG must be able to reassure the patient that this is without additional risk compared with open surgery.

Prophylactic mastectomy

Prophylactic mastectomy is not routinely recommended but may be a reasonable option for some women. Literature about prophylactic mastectomy in HDGC is scarce, and it is reasonable to consider prophylactic mastectomy on a case-by-case basis taking into account the family pedigree. National guidelines for

high-risk women should be followed with respect to chemoprevention using selective oestrogen receptor modulators or aromatase inhibitors.

HISTOPATHOLOGY

Histopathology of biopsies from individuals suspected for HDGC

Biopsies of individuals with a family history of DGC that are endoscopically screened by the Cambridge protocol should be examined thoroughly. The biopsies should be stained for H&E at three levels and periodic acid–Schiff–diastase (PAS-D) as standard. All lesions should be recorded. Biopsies from potential carriers of a *CDH1* mutation need careful evaluation by a pathologist with experience in the pathology of HDGC. In mutation carriers, the detection of specific lesions may support a decision towards prophylactic gastrectomy. In individuals without known gene defects, the detection of such lesions may be used as an additional argument that the patient is at high risk of developing cancer and that a prophylactic gastrectomy should be discussed. The specific lesions in HDGC are tiny *foci* of typical signet ring cells sometimes admixed with a low number of smaller atypical cells in the lamina propria without infiltration beneath the muscularis mucosa (see [figure 3A–D](#)). The two preinvasive lesions of SRCC are (1) *in situ* SRCC, corresponding to the presence of signet ring cells with hyperchromatic and depolarised nuclei within the basal membrane of a gland replacing the normal cells of the gland; and (2) pagetoid spread of a row of signet ring cells below the preserved epithelium of glands and foveolae, and also within the basal membrane (see [figure 4A, B](#)).⁸⁴

Criteria for the identification of signet ring cell lesions should be strictly followed in order to diminish the risk of overdiagnosing non-specific changes and to distinguish them from lesions that mimic SRCC or precursor lesions. In the gastrointestinal tract, various benign ‘signet cell-like changes’ may mimic SRCC (see online supplementary figure S1).^{85–88} Therefore, confirmation of focal signet ring cell lesions in the stomach by a histopathologist with experience in this area is strongly recommended.

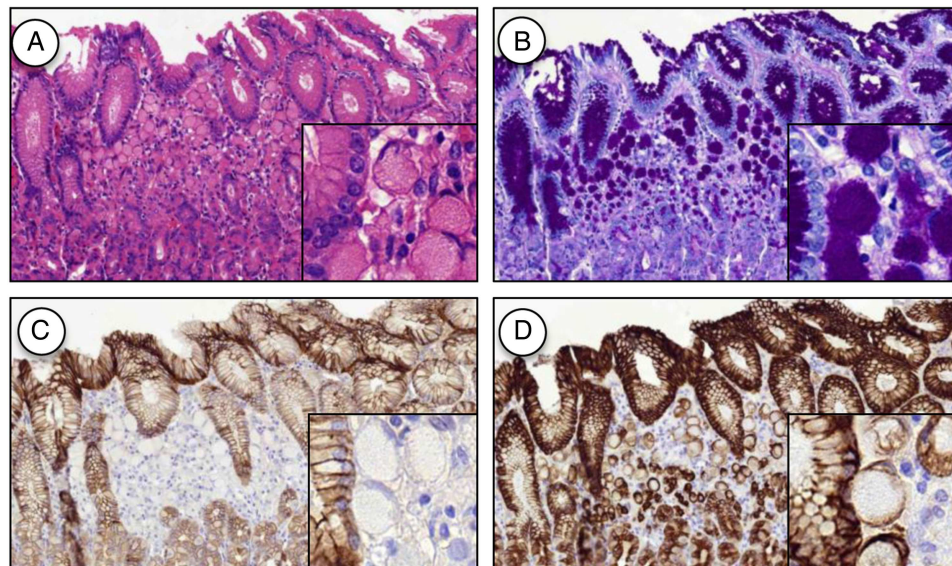
Histopathology of prophylactic gastrectomy: classification of microscopic foci and determination of significance

Macroscopic examination and sampling of prophylactic gastrectomies should follow specific protocols that can be found in online supplementary protocol 2. The histological examination should be made using a checklist, focusing on the items listed in [table 1](#).

Nearly all gastrectomies (data from >100 PTG cases published) exhibit tiny mucosal *foci* of SRCC or *in situ* signet ring cells, although sometimes these were only discovered after careful review by a pathologist with experience in this field.^{25–36–39–59–64–77–91–92} There are numerous T1a carcinoma *foci* but only a low number of *in situ* carcinoma lesions, suggesting that invasion of the lamina propria by signet ring cells may occur without a morphologically detectable *in situ* carcinoma.^{63–93} It is essential that the location of biopsies within gastrectomy specimens is specifically reported to learn more about the distribution of early HDGC in the stomach.

Surgical margin status must confirm that there is no residual gastric mucosa and tumour at the margins. However, oesophageal cardiac-type glands (presumably normal constituents) are diffusely scattered in the lamina propria through all levels of the oesophagus. The risk to develop SRCC in these glands is unknown but has not been reported.⁹⁴ Also, metaplastic and heterotopic gastric mucosa can be seen elsewhere in the gastrointestinal tract. In a prophylactic gastrectomy series of 19

Figure 3 Mucosal signet ring cell carcinoma (pT1a) H&E (A), periodic acid–Schiff–diastase (B), E-cadherin (C) and cytokeratin staining (D) (original magnifications $\times 200$).



patients, three cases (16%) showed heterotopic gastric mucosa in the duodenum, and mucosal SRCC was observed in one of them (RT van der Kaaij *et al*, unpublished data, 2014).

Background changes in the gastric mucosa of prophylactic gastrectomy specimens encompass mild chronic gastritis, sometimes displaying the features of lymphocytic gastritis. Foveolar hyperplasia and tufting of surface epithelium, focally with globoid change, is also a frequent finding and, in some areas, vacuolisation of surface epithelium is very striking; however, this does not seem to be a specific finding.^{28 36 63 93} Intestinal metaplasia and *H. pylori* infection are absent in most prophylactic gastrectomies described to date.

Histopathology: advanced HDGC

Like sporadic DGC, advanced HDGC predominantly presents as *limitis plastica* with diffuse infiltration of the gastric wall. Histology can show mainly or exclusively signet ring cells; however, more often these tumours are composed of a pleomorphic neoplastic infiltrate with a small subset of or without classic signet ring cells (poorly cohesive carcinoma). In a minority of cases, tumour cells are arranged in small aggregates, sometimes rosettes or glandular-like structures. A component of extracellular mucin may also be present, in which the neoplastic cells float. Although there are no specific morphological

characteristics indicating the hereditary nature of the tumour, *in situ* lesions and pagetoid spread of signet ring cells in the surrounding normal mucosa are important clues to the diagnosis.

Review of gastric specimens of 103 HDGC families without germline *CDH1* mutations showed a similar morphology compared with the advanced germline *CDH1* mutation-related carcinomas (van der Post *et al*, unpublished data, 2014). Typical *in situ* lesions or pagetoid spread of signet ring cells, however, were not detected in these patients.

Histopathology: LBC

The association of LBC and germline *CDH1* mutations was first reported by Keller *et al*.⁹⁵ Large series of the histopathological findings in (prophylactic) mastectomies of *CDH1* mutation carriers are not reported. Kluijdt *et al*³⁶ described bilateral wide-spread *foci* of lobular carcinoma *in situ* in prophylactic mastectomies of two female patients with germline *CDH1* mutations. There are no unique histopathological findings reported in (prophylactic) mastectomy specimens that can be used to distinguish these (pre-)malignancies from sporadic LBCs. However, only few patients opt for prophylactic mastectomy and these mastectomy specimens are generally not fully embedded and examined.

Figure 4 Precursor signet ring cell lesions: pagetoid spreading of signet ring cells (A) and *in situ* signet ring cell carcinoma (B) (H&E, original magnifications $\times 400$).

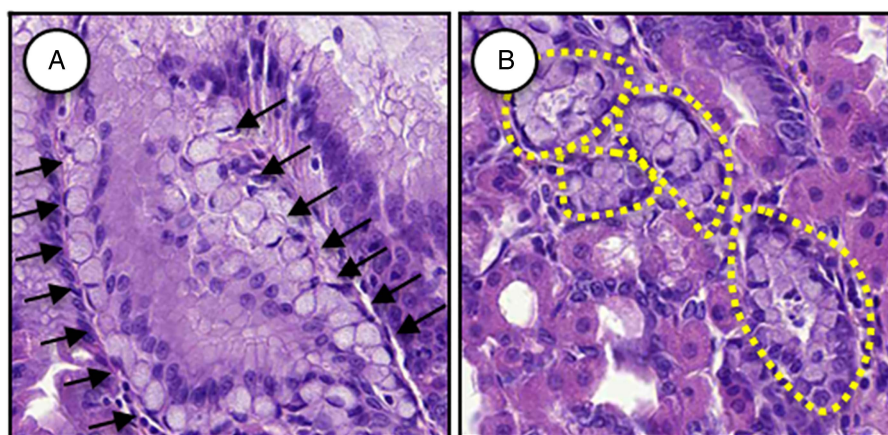


Table 1 Checklist for reporting of prophylactic gastrectomy specimens

1. Features of \geq pT1b carcinoma(s)	Growth pattern (diffuse infiltration vs localised tumour) Anatomic location (cardia, fundus, body, transitional zone, antrum) Measurements Histological type according to WHO ⁸⁹ and Laurén's ⁹⁰ classifications Lymphatic, venous and neural invasion (present or absent) Tumour, node, metastases stage
2. Features of intramucosal precursor lesions and pT1a SRCC	Number of lesions Anatomic location (cardia, fundus, body, transitional zone, antrum) Measurements Aggressive features: pleomorphism, loss of mucin, spindle cells, small cells, mitoses Stromal reaction related to lesions: desmoplasia; lymphocytic, eosinophilic or granulomatous inflammatory reaction Surgical margin status (proximal oesophageal, distal duodenal mucosa, including donuts), to confirm there is no residual gastric mucosa and no tumour at margins. Lymph node status
3. Non-neoplastic mucosa: changes more commonly seen in this condition	Tufting/hyperplastic mucosal changes Surface epithelial vacuolisation Globoid change
4. Other findings in surrounding mucosa	Inflammation (acute, chronic, erosion, ulceration) <i>Helicobacter pylori</i> Intraepithelial lymphocytes Lymphoid infiltrates Glandular atrophy Intestinal metaplasia Adenomatous dysplasia

SRCC, signet ring cell carcinoma.

Use of immunohistochemical stains

Widely invasive DGC may be easily detected on H&E sections. The use of histochemical stains for neutral mucins, such as PAS-D, is useful for the detection or confirmation of tiny intramucosal carcinomas in which the neoplastic cells are dispersed among preserved foveolae and glands. This should be performed routinely in the examination of gastric biopsies taken during endoscopy and in gastrectomies from patients with HDGC.⁹⁶ A cytokeratin stain can help to confirm the epithelial nature of the signet ring cells if there is any doubt. E-cadherin immunoexpression can be reduced or absent in early gastric carcinomas, contrasting with the normal membranous E-cadherin expression in adjacent non-neoplastic mucosa.^{39 40 97} In pagetoid spread and *in situ* carcinomas, E-cadherin immunoexpression can also be reduced or absent.⁶³ However, E-cadherin expression is not always reduced or absent; this depends on the mutation type and specific mechanisms of inactivation of the wild-type allele.⁴⁰ In DGC of patients without germline *CDH1* mutations, the expression pattern of E-cadherin is also often reduced or absent. Therefore, E-cadherin staining should not be used as a pre-screening method to select patients eligible for germline *CDH1* mutation analysis.

In patients that present with both LBC and DGC, either synchronously or metachronously, a metastasis should be considered. Two primary tumours may be indicative for a hereditary background, but this is not always the underlying reason. Metastases from these tumours are often morphologically indistinguishable.⁹⁸ Breast-associated immunomarkers are ER, BRST-2 and/or mammaglobin, while CK20 and HNF4A are suggestive for GC.^{99 100}

Centres of excellence for pathological diagnosis

The pathology of HDGC is unique but not easy to recognise. Experience in this pathology is needed to provide high-quality diagnosis, both in biopsies and in resection specimens. In order to increase the experience of pathologists and the accuracy of the diagnosis, especially of precursor lesions of HDGC, it

would be useful to build a virtual bank of the different types of lesions observed in the setting of HDGC. Furthermore, the working group agreed that the use of (scanned) slides to be submitted for evaluation by experienced pathologists in the field should be seriously considered.

It is pivotal to examine the full gastrectomy and full mastectomy specimens of *CDH1* mutation carriers to determine the stage of cancer and additionally to better understand the phenotype and biology of this disease. Experience in the examination of prophylactic gastrectomies for HDGC is quite limited in most pathology departments due to the rarity of these surgical specimens. Additionally, the routine workload of most centres is incompatible with performing the detailed examination of hundreds of sections typically obtained after totally embedding these stomachs. In the event of a lack of pathologist experience in dealing with these cases, or restricted time available due to the pathologist's workload and laboratory resources, the entire formalin-fixed gastrectomy or mastectomy specimen can be sent to an experienced pathology laboratory. An alternative option is to totally embed the stomach or breast, perform H&E and PAS-D stain on all blocks and send the slides and blocks to an experienced centre for specialist pathology reporting. If these alternative strategies are not feasible, and it is not possible to totally embed the gastric or breast specimen, this should be communicated to clinicians and the patient. Furthermore, in the event of not finding *foci* of SRCC, the gastrectomy should not be reported as negative for carcinoma, but as 'no carcinoma found in xx% of mucosa examined'.

Future research on molecular pathogenesis

A critical question that remains unanswered is how long early lesions of HDGC can remain indolent until there is emergence of clinical disease that may be rapidly progressive and lethal. Continuing collection of data from patients who opt for endoscopic surveillance is essential to help answer this question. Also, a thorough analysis of the mechanisms responsible for the

Table 2 Postgastrectomy symptoms and treatment options

Symptoms	Treatment
Early dumping (15–30 min after eating)	Modification of diet and eating habits
Late dumping (1.5–3 h after eating)	Modification of diet and eating habits
Lactose intolerance	Diet modifications, supplementation with lactase enzymes
Fat malabsorption	Pancreatic enzyme replacement may be necessary, especially fats and fat-soluble vitamins such as vitamin D ¹⁰²
Small bowel bacterial overgrowth/blind loop syndrome	Antibiotics, sometimes surgery
Dysphagia and anastomotic strictures	Modification of eating habits (more deliberate mastication and smaller bites) Upper endoscopy with balloon dilatation to widen the stricture Sometimes postsurgical intervention
Changing response to usage of alcohol	Alertness physician and patient
Absorption of medication can be affected	Alertness physician and patient
Monitoring deficiencies of iron, vitamin B12, folate and trace elements	Supplementing of vitamin B12 and/or folate. ^{103–105} Intravenous iron infusions may become necessary if individuals are unable to tolerate iron orally, unable to absorb sufficient iron through the gut, or require rapid increase in iron levels to avoid other health complications or a blood transfusion
Hypocalcaemia, osteoporosis, osteopenia and osteomalacia	Calcium and vitamin D are required to reduce risk of bone disease. When levels cannot be maintained through consumption of calcium-rich foods, supplements such as vitamin D, calcium carbonate, calcium phosphate and calcium citrate can be used. Calcium citrate does not require acid for absorption, making it a strong choice of supplement ¹⁰⁶
Bile reflux (due to the absence of the gastroesophageal valve)	Type of surgical procedure (appropriate length of the Roux limb) reduces the occurrence of bile reflux Use of wedge pillow for sleeping Limiting oral intake to 2–3 h before going to bed

second hit inactivation of *CDH1* in the very early lesions of HDGC is necessary to define strategies for chemoprevention.

The molecular background of patients with HDGC and without *CDH1* mutations remains to be clarified, including any specific morphological features of GC in the setting of other hereditary cancer syndromes. Majewski *et al*²³ reported a family with a germline *CTNNA1* mutation presenting with multiple DGC cases and intramucosal signet ring cells. Immunohistochemistry of α -E-catenin showed absent staining in the signet ring cells, while tumours from 10 other HDGC pedigrees without *CDH1* and *CTNNA1* mutation stained positively. Two additional families have recently been described, one of which shared an immunohistochemical phenotype with loss of α -catenin and cytoplasmic rather than membranous E-cadherin.²⁴ This finding suggests either that the pathogenicity of *CTNNA1* mutations may be mediated through loss of E-cadherin function or that the cancer-initiating potential of *CDH1* mutations is imparted through α -catenin-associated pathways. More research is needed to understand the role and mutation detection rate of *CTNNA1* mutations.

POSTSURGICAL CARE AND NUTRITION

Postoperative care

The psychological, physiological and metabolic impact of a total gastrectomy should not be underestimated. The physical impact of a gastrectomy is difficult to predict for any individual, but there is an expectation that most patients will return to a full and active life after their operation. Reassuringly, global quality-of-life scores recover to presurgery levels at around 12 months postoperatively; however, problems with eating, abdominal pain and reduced body image persist beyond this time.¹⁰¹

Enhanced postgastrectomy recovery programmes are now well established. These programmes may include preoperative carbohydrate loading, preferably the avoidance of a nasogastric tube or abdominal drain, early mobilisation with good analgesia (epidural or local anaesthetic wound catheters), resume oral intake within 3–5 days and discharge from hospital within 7–10 days. In addition, patients often require considerable support during

the first 12 months after surgery. Implementation of this support will depend upon the local healthcare services and the distances required for patients to attend the centre. Specialist nurses and dietitians should maintain regular contact by telephone and the use of modern video-conferencing should be encouraged. Collection of clinical data in a national registry allows patients, if they choose, to contribute to HDGC syndrome research.

There are different issues related to mastectomy (prophylactic or otherwise), a second major surgery with a significant recovery period. Mastectomy has a different impact concerning self-image, self-esteem, physical appearance and loss of feminine identity. Therefore, a team including a dietician, physiotherapist and psychologist should be available for optimal physical and socio-emotional recovery during postoperative care of both gastrectomy and mastectomy.

Nutrition

The main adjustments with regard to diet and nutrition postgastrectomy have to do with (1) maintaining weight; (2) ensuring adequate fluid, nutritional and caloric intake; and (3) behavioural modifications surrounding eating. Experienced dietitians focus on nutritional problems and strategies for maintaining weight after surgery, while patients focus on lifestyle changes. Patients are often discouraged by weight loss. The median weight loss 1 year postsurgery is 10 kg.¹⁰¹ This means that patients who are underweight preoperatively or who have a history of eating disorders need very careful counselling and support. Anatomical changes can make the act of eating difficult, and patients may become disappointed by these hurdles. This can further complicate weight management with the psychological burden of eating. In the early stages of recovery, intentional eating, drinking, management of symptoms and resting can quickly become draining. It is important for patients to have realistic expectations for their progress and improvement over time. Common postsurgical symptoms, risks and treatment options are listed in [table 2](#).

Following a prophylactic gastrectomy, patients initially have to eat frequent small meals. Eating too much and/or too quickly will cause abdominal pain. Dumping syndrome is a group of

symptoms commonly experienced as a result of altered gut anatomy following gastrectomy, caused by rapid entry of food into the small intestine at an earlier stage of digestion. This leads to a shift of fluid from the bloodstream to the small intestine to aid in digestion, and may lead to cardiovascular and abdominal symptoms. Late dumping is caused by a rapid rise and subsequent decrease in blood sugar levels by delayed insulin secretion. After gastrectomy, food moves rapidly and directly into the small intestine, where it digests faster. Also, the pancreas produces more insulin in a short time. This excessive insulin production allows for a rapid decrease in the blood sugar.¹⁰⁷ These symptoms may be more common in the immediate postoperative period and often subside over time. Dumping syndrome can be minimised or eliminated through dietary choices and modified eating habits; adequate adjustments are unique from patient to patient. With time, however, these symptoms tend to improve.¹⁰¹ Other problems may include lactose intolerance, steatorrhoea, small bowel bacterial overgrowth, anastomotic strictures and postprandial fullness.^{108–109} Patients should be warned that their tolerance to alcohol will reduce after gastrectomy and that absorption of some medications can be affected.

As a result of malabsorption, patients with postgastrectomy are at risk for nutritional deficiencies. Monitoring of nutritional levels in postsurgical follow-up is essential as deficiencies increase risk for other symptoms and health concerns. All patients require lifelong vitamin B12 supplementation (either oral, subcutaneous or intramuscularly) to correct identified deficiencies, and close monitoring for conditions such as iron deficiency, anaemia, hypocalcaemia, osteoporosis and trace element deficiencies. Many dietitians recommend patients to take a daily multivitamin preparation with iron; however, it remains important to monitor iron levels since absorption will change after gastrectomy. While these are the most commonly reported symptoms, attention should be paid to any significant and prolonged changes, such as hair loss or extreme fatigue. These symptoms and changes away from the patient's baseline may be indicative of nutritional deficiencies, which may need to be identified and treated.

Patients should consult a dietician prior to surgery as an awareness of baseline nutritional status and dietary habits will benefit the patient in postsurgical nutrition, diet and weight management. While there are some basic dietary principles that apply to most patients with gastrectomy, there are no absolute rules. Each patient's recovery is unique, from food and quantity tolerances, to comfortable eating habits. Variability is observed between patients, but also for individual patients during the course of their recovery. The most notable changes occur within the first year; however, deficiencies should be carefully followed since they can also develop over years. Patients are encouraged to continue to experiment and discover what is best suited to their needs and tolerances.

Future research: nutrition postgastrectomy

Further studies are needed to elucidate the relationship in patients with postgastrectomy between diet, nutrition, drug absorption, changes in body composition and the direct impact on quality of life, both in the short term and long term.

PATIENT ADVOCACY AND THE NEXT STEPS IN PATIENT CARE AND HDGC RESEARCH

In patients' experience, the process that HDGC families go through to find local medical care providers with experience with HDGC is frustrating. Faced with the lack of universal

expertise, patients and advocacy groups have started working together to locate local care providers with expertise or facilitate connecting local care providers with experts at globally recognised multidisciplinary centres of excellence. Medical expertise in HDGC has to continue to grow, and this expertise has to be communicated within the medical community, such that support structures for patients with HDGC will become more easily accessible and established within existing medical systems.

Given the rare nature of HDGC, there is currently no singular global expertise portal for genetic counselling, psychosocial support, gastroenterology, surgery, pathology and postsurgery follow-up for HDGC families. The geography of existing expertise centres does not nearly cover broadly enough the areas where these resources are needed. With the emerging trend towards telemedicine, and the development of various patient advocacy groups, notably 'No Stomach For Cancer',¹¹⁰ 'DeGregorio Family Foundation for Stomach and Esophageal Cancer Research',¹¹¹ in the USA and 'Stichting CDH1',¹¹² in the Netherlands, we see the potential for collaboration between patients, medical professionals and patient advocacy groups to the end of empowering patients by directing and connecting them with the appropriate resources and expert opinions.

Author affiliations

¹Department of Pathology, Radboud University Medical Center, Nijmegen, The Netherlands

²Department of Human Genetics, Radboud University Medical Center, Nijmegen, The Netherlands

³Institute of Molecular Pathology and Immunology of the University of Porto (IPATIMUP), Porto, Portugal

⁴Instituto de Investigação e Inovação em Saúde, Universidade do Porto, Porto, Portugal

⁵Department of Pathology and Oncology, Medical Faculty of the University of Porto, Porto, Portugal

⁶Centro Hospitalar São João, Porto, Portugal

⁷Department of Biochemistry, University of Otago, Dunedin, New Zealand

⁸British Columbia Cancer Agency, Vancouver, British Columbia, Canada

⁹Department of Oncology, University of Cambridge, Cambridge, UK

¹⁰No Stomach For Cancer, Inc., Madison, Wisconsin, USA

¹¹Department of Oesophago-Gastric Surgery, Addenbrooke's Hospital, Cambridge, UK

¹²Department of Medical Genetics, University Medical Centre Utrecht, Utrecht, The Netherlands

¹³Department of Surgical Gastroenterology, Rigshospitalet, University of Copenhagen, Copenhagen, Denmark

¹⁴Department of Medical Oncology, Institut Gustave Roussy, Villejuif, France

¹⁵Department of Gastroenterology, Radboud University Medical Center, Nijmegen, The Netherlands

¹⁶Vinery Lane Surgery, Whangarei, New Zealand

¹⁷Division of Psychosocial Research and Epidemiology/Family Cancer Clinic, The Netherlands Cancer Institute, Amsterdam, The Netherlands

¹⁸Sir Peter MacCallum Department of Oncology, Peter MacCallum Cancer Centre, Melbourne, Victoria, Australia

¹⁹Department of Medicine, Royal Melbourne Hospital, University of Melbourne, Melbourne, Victoria, Australia

²⁰Department of Gastroenterology and Hepatology, Netherlands Cancer Institute/Antoni van Leeuwenhoek Hospital, Amsterdam, The Netherlands

²¹Department of Surgery, Memorial Sloan-Kettering Cancer Center, New York, New York, USA

²²The DeGregorio Family Foundation for Stomach and Esophageal Cancer Research, Pleasantville, New York, USA

²³Division of Oncology, Stanford University School of Medicine, Stanford, California, USA

²⁴Netherlands Cancer Institute/Antoni van Leeuwenhoek Hospital, Amsterdam, The Netherlands

²⁵Scientific Institute for Quality of Healthcare, Radboud University Medical Center, Nijmegen, The Netherlands

²⁶Division of Surgical Research, University of Zurich, Zurich, Suisse

²⁷Department of Medical Genetics, University of British Columbia, Vancouver, British Columbia, Canada

²⁸Institute of Pathology, Technische Universität, Munich, Germany

²⁹Department of Histopathology, Cambridge University Hospitals NHS Trust, Cambridge, UK

Review

³⁰NIHR Biomedical Research Unit, Nottingham Digestive Diseases Centre, Queens Medical Centre campus, Nottingham University Hospitals NHS Trust, Nottingham, UK
³¹Patient representative, The Netherlands
³²Department of Oncology, Familial Gastric Cancer Registry, Cambridge University Hospital, Cambridge, UK
³³Department of General Surgery and Surgical Oncology, University of Siena, Siena, Italy
³⁴Department of Surgical Research, Technical University Dresden, Dresden, Germany
³⁵Cambridge University Hospital, Cambridge, UK
³⁶Stichting CDH1, The Netherlands
³⁷Department of Medical Genetics, University of Cambridge, Cambridge, UK
³⁸Department of Pathology, VU University Medical Center, Amsterdam, The Netherlands
³⁹Department of Surgical Oncology, University Medical Centre Utrecht, Utrecht, The Netherlands
⁴⁰Department of Surgery, Netherlands Cancer Institute/Antoni van Leeuwenhoek Hospital, Amsterdam, The Netherlands
⁴¹Radboud University Medical Center, Nijmegen, The Netherlands
⁴²Cambridge NIHR Biomedical Research Centre, University of Cambridge NHS Foundation Trust
⁴³MRC Cancer Unit, Hutchison/MRC Research Centre, Cambridge, UK
⁴⁴Department Gastroenterology, Cambridge University Hospitals, UK

Correction notice This article has been corrected since it published Online First. The order of the first two authors has been corrected.

Acknowledgements We are very grateful to Bert Siebers from the Department of Pathology of the Radboudumc for excellent assistance in the organisation of the consensus meeting. We kindly acknowledge Patricia Oliveira, the Bioinformatician that helped collecting the information for the CDH1 database, and the support of the LOVD-team (Leiden University Medical Center, Leiden, the Netherlands) for setting up, maintaining and hosting the CDH1 database.

Contributors RSvdP and IPV contributed equally. Concept and design: RSvdP, IPV, JHVk and RCF. Acquisition of data, analysis and interpretation of data, critical revision of the manuscript for important intellectual content, final approval of manuscript: all authors. Drafting of the manuscript: RSvdP, IPV, FC, PG, DH, NH, JHVk, RCF with input of all authors.

Funding The 8th (Cambridge, Vancouver, Porto, Munich, Siena, Seoul, Cambridge and Nijmegen) workshop of the International Gastric Cancer Linkage Consortium was made possible by generous sponsorship from the Royal Netherlands Academy of Arts and Sciences (KNAW), the Netherlands Organisation for Health Research and Development (ZonMW) and the European Society of Pathology.

Competing interests None.

Provenance and peer review Not commissioned; externally peer reviewed.

Open Access This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

REFERENCES

- 1 Ferlay J. GLOBOCAN 2012 v1.0, cancer incidence and mortality worldwide. IARC CancerBase 11.
- 2 Oliveira C, Ferreira P, Nabais S, Campos L, Ferreira A, Cirnes L, Alves CC, Veiga I, Fragoso M, Regateiro F, Dias LM, Moreira H, Suriano G, Machado JC, Lopes C, Castedo S, Carneiro F, Seruca R. E-Cadherin (CDH1) and p53 rather than SMAD4 and Caspase-10 germline mutations contribute to genetic predisposition in Portuguese gastric cancer patients. *Eur J Cancer* 2004;40:1897–903.
- 3 Keller G, Vogelsang H, Becker I, Plaschke S, Ott K, Suriano G, Mateus AR, Seruca R, Biedermann K, Huntsman D, Doring C, Holinski-Feder E, Neutzing A, Siewert JR, Hoffer H. Germline mutations of the E-cadherin(CDH1) and TP53 genes, rather than of RUNX3 and HPP1, contribute to genetic predisposition in German gastric cancer patients. *J Med Genet* 2004;41:e89.
- 4 Masciari S, Dewanwala A, Stoffel EM, Lauwers GY, Zheng H, Achatz MI, Riegert-Johnson D, Foretova L, Silva EM, Digiani L, Verselis SJ, Schneider K, Li FP, Fraumeni J, Garber JE, Syngal S. Gastric cancer in individuals with Li-Fraumeni syndrome. *Genet Med* 2011;13:651–7.
- 5 Park YJ, Shin KH, Park JG. Risk of gastric cancer in hereditary nonpolyposis colorectal cancer in Korea. *Clin Cancer Res* 2000;6:2994–8.
- 6 Capelle LG, Van Grieken NC, Lingsma HF, Steyerberg EW, Klokman WJ, Bruno MJ, Vasen HF, Kuipers EJ. Risk and epidemiological time trends of gastric cancer in Lynch syndrome carriers in the Netherlands. *Gastroenterology* 2010;138:487–92.
- 7 Sereno M, Aguayo C, Guillen Ponce C, Gomez-Raposo C, Zambrana F, Gomez-Lopez M, Casado E. Gastric tumours in hereditary cancer syndromes: clinical features, molecular biology and strategies for prevention. *Clin Transl Oncol* 2011;13:599–610.
- 8 van Lier MG, Westerman AM, Wagner A, Looman CW, Wilson JH, de Rooij FW, Lemmens VE, Kuipers EJ, Mathus-Vliegen EM, van Leerdam ME. High cancer risk and increased mortality in patients with Peutz-Jeghers syndrome. *Gut* 2011;60:141–7.
- 9 Giardiello FM, Brensinger JD, Tersmette AC, Goodman SN, Petersen GM, Booker SV, Cruz-Correa M, Offerhaus JA. Very high risk of cancer in familial Peutz-Jeghers syndrome. *Gastroenterology* 2000;119:1447–53.
- 10 Giardiello FM, Trimbath JD. Peutz-Jeghers syndrome and management recommendations. *Clin Gastroenterol Hepatol* 2006;4:408–15.
- 11 Jakubowska A, Nej K, Huzarski T, Scott RJ, Lubinski J. BRCA2 gene mutations in families with aggregations of breast and stomach cancers. *Br J Cancer* 2002;87:888–91.
- 12 Friedenson B. BRCA1 and BRCA2 pathways and the risk of cancers other than breast or ovarian. *MedGenMed* 2005;7:60.
- 13 Win AK, Hopper JL, Jenkins MA. Association between monoallelic MUTYH mutation and colorectal cancer risk: a meta-regression analysis. *Fam Cancer* 2011;10:1–9.
- 14 Iwama T, Mishima Y, Utsunomiya J. The impact of familial adenomatous polyposis on the tumorigenesis and mortality at the several organs—its rational treatment. *Ann Surg* 1993;217:101–8.
- 15 Park SY, Ryu JK, Park JH, Yoon H, Kim JY, Yoon YB, Park JG, Lee SH, Kang SB, Park JW, Oh JH. Prevalence of gastric and duodenal polyps and risk factors for duodenal neoplasm in Korean patients with familial adenomatous polyposis. *Gut Liver* 2011;5:46–51.
- 16 Lynch HT, Snyder C, Davies JM, Lanspa S, Lynch J, Gatalica Z, Graeve V, Foster J. FAP, gastric cancer, and genetic counseling featuring children and young adults: a family study and review. *Fam Cancer* 2010;9:581–8.
- 17 Howe JR, Mitros FA, Summers RW. The risk of gastrointestinal carcinoma in familial juvenile polyposis. *Ann Surg Oncol* 1998;5:751–6.
- 18 Pollock J, Welsh JS. Clinical cancer genetics: Part I: Gastrointestinal. *Am J Clin Oncol* 2011;34:332–6.
- 19 Stanich PP, Francis DL, Sweetser S. The spectrum of findings in Cowden syndrome. *Clin Gastroenterol Hepatol* 2011;9:e2–3.
- 20 Guilford P, Hopkins J, Harraway J, McLeod M, McLeod N, Harawira P, Taite H, Scoular R, Miller A, Reeve AE. E-cadherin germline mutations in familial gastric cancer. *Nature* 1998;392:402–5.
- 21 Caldas C, Carneiro F, Lynch HT, Yokota J, Wiesner GL, Powell SM, Lewis FR, Huntsman DG, Pharoah PD, Jankowski JA, MacLeod P, Vogelsang H, Keller G, Park KG, Richards FM, Maher ER, Gayther SA, Oliveira C, Grehan N, Wight D, Seruca R, Roviello F, Ponder BA, Jackson CE. Familial gastric cancer: overview and guidelines for management. *J Med Genet* 1999;36:873–80.
- 22 Fitzgerald RC, Hardwick R, Huntsman D, Carneiro F, Guilford P, Blair V, Chung DC, Norton J, Ragnauth K, Van Krieken JH, Derryhouse S, Caldas C, International Gastric Cancer Linkage C. Hereditary diffuse gastric cancer: updated consensus guidelines for clinical management and directions for future research. *J Med Genet* 2010;47:436–44.
- 23 Majewski IJ, Kluij I, Cats A, Scerri TS, de Jong D, Kluij RJ, Hansford S, Hogervorst FB, Bosma AJ, Hofland I, Winter M, Huntsman D, Jonkers J, Bahlo M, Bernards R. An alpha-E-catenin (CTNNA1) mutation in hereditary diffuse gastric cancer. *J Pathol* 2013;229:621–9.
- 24 Hansford S, Kaurah P, Li-Chang H, Woo M, Senz J, Pinheiro H, Schrader KA, Schaeffer DF, Shumansky K, Zogopoulos G, Almeida Santos T, Claro I, Carvalho J, Nielsen C, Padilla S, Lum A, Talhouk A, Baker-Lange K, Richardson S, Lewis I, Lindor NM, Pennell E, MacMillan A, Fernandez B, Keller G, Lynch H, Shah SP, Guilford P, Gallinger S, Corso G, Roviello F, Caldas C, Oliveria C, Pharoah PDP, Huntsman DG. Hereditary diffuse gastric cancer syndrome: CDH1 mutations and beyond. *JAMA Oncol* 2015. doi:10.1001/jamaoncol.2014.168
- 25 Wickremaratne T, Lee CH, Kirk J, Charlton A, Thomas G, Gaskin KJ. Prophylactic gastrectomy in a 16-year-old. *Eur J Gastroenterol Hepatol* 2014;26:353–6.
- 26 Pharoah PD, Guilford P, Caldas C, International Gastric Cancer Linkage C. Incidence of gastric cancer and breast cancer in CDH1 (E-cadherin) mutation carriers from hereditary diffuse gastric cancer families. *Gastroenterology* 2001;121:1348–53.
- 27 Kaurah P, MacMillan A, Boyd N, Senz J, De Luca A, Chun N, Suriano G, Zaor S, Van Manen L, Gilpin C, Nikkel S, Connolly-Wilson M, Weissman S, Rubinstein WS, Sebold C, Greenstein R, Stroop J, Yim D, Panzini B, McKinnon W, Greenblatt M, Wirtzfeld D, Fontaine D, Coit D, Yoon S, Chung D, Lauwers G, Pizzuti A, Vaccaro C, Redal MA, Oliveira C, Tischkowitz M, Olshwang S, Gallinger S, Lynch H, Green J, Ford J, Pharoah P, Fernandez B, Huntsman D. Founder and recurrent CDH1 mutations in families with hereditary diffuse gastric cancer. *JAMA* 2007;297:2360–72.
- 28 Oliveira C, Seruca R, Carneiro F. Genetics, pathology, and clinics of familial gastric cancer. *Int J Surg Pathol* 2006;14:21–33.
- 29 Benusiglio PR, Malka D, Rouleau E, De Pauw A, Buecher B, Nogues C, Fourme E, Colas C, Coulet F, Warcoin M, Grandjouan S, Sezeur A, Laurent-Puig P, Moliere D, Tlemsani C, Di Maria M, Byrde V, Delaloge S, Blayau M, Caron O. CDH1 germline

- mutations and the hereditary diffuse gastric and lobular breast cancer syndrome: a multicentre study. *J Med Genet* 2013;50:486–9.
- 30 Kim S, Chung JW, Jeong TD, Park YS, Lee JH, Ahn JY, Kim do H, Choi KD, Lee W, Song HJ, Lee GH, Chun S, Jung HY, Min WK, Kim JH. Searching for E-cadherin gene mutations in early onset diffuse gastric cancer and hereditary diffuse gastric cancer in Korean patients. *Fam Cancer* 2013;12:503–7.
 - 31 Choi HJ, Ki CS, Suh SP, Kim JW. Presymptomatic identification of CDH1 germline mutation in a healthy Korean individual with family history of gastric cancer. *Ann Lab Med* 2014;34:386–9.
 - 32 Yamada H, Shinmura K, Ito H, Kasami M, Sasaki N, Shima H, Ikeda M, Tao H, Goto M, Ozawa T, Tsuneyoshi T, Tanioka F, Sugimura H. Germline alterations in the CDH1 gene in familial gastric cancer in the Japanese population. *Cancer Sci* 2011;102:1782–8.
 - 33 Yamada M, Fukagawa T, Nakajima T, Asada K, Sekine S, Yamashita S, Okochi-Takada E, Taniguchi H, Kushima R, Oda I, Saito Y, Ushijima T, Katai H. Hereditary diffuse gastric cancer in a Japanese family with a large deletion involving CDH1. *Gastric Cancer* 2014;17:750–6.
 - 34 Petridis C, Shinomiya I, Kohut K, Gorman P, Caneppele M, Shah V, Troy M, Pinder SE, Hanby A, Tomlinson I, Trembath RC, Roylance R, Simpson MA, Sawyer EJ. Germline CDH1 mutations in bilateral lobular carcinoma in situ. *Br J Cancer* 2014;110:1053–7.
 - 35 Frebourg T, Oliveira C, Hochain P, Karam R, Manouvrier S, Graziadio C, Vekemans M, Hartmann A, Baert-Desurmont S, Alexandre C, Lejeune Dumoulin S, Marroni C, Martin C, Castedo S, Lovett M, Winston J, Machado JC, Attie T, Jabs EW, Cai J, Pellerin P, Triboulet JP, Scotte M, Le Pessot F, Hedouin A, Carneiro F, Blayau M, Seruca R. Cleft lip/palate and CDH1/E-cadherin mutations in families with hereditary diffuse gastric cancer. *J Med Genet* 2006;43:138–42.
 - 36 Kluijlt I, Siemerink EJ, Ausems MG, van Os TA, de Jong D, Simoes-Correia J, van Krieken JH, Ligtgenberg MJ, Figueiredo J, van Riel E, Sijmons RH, Plukker JT, van Hillegersberg R, Dekker E, Oliveira C, Cats A, Hoogerbrugge N, Dutch Working Group on Hereditary Gastric C. CDH1-related hereditary diffuse gastric cancer syndrome: clinical variations and implications for counseling. *Int J Cancer* 2012;131:367–76.
 - 37 Oliveira C, Senz J, Kaurah P, Pinheiro H, Sanges R, Haegert A, Corso G, Schouten J, Fitzgerald R, Vogelsang H, Keller G, Dwerryhouse S, Grimmer D, Chin SF, Yang HK, Jackson CE, Seruca R, Roviello F, Stupka E, Caldas C, Huntsman D. Germline CDH1 deletions in hereditary diffuse gastric cancer families. *Hum Mol Genet* 2009;18:1545–55.
 - 38 Guilford P, Humar B, Blair V. Hereditary diffuse gastric cancer: translation of CDH1 germline mutations into clinical practice. *Gastric Cancer* 2010;13:1–10.
 - 39 Barber M, Murrell A, Ito Y, Maia AT, Hyland S, Oliveira C, Save V, Carneiro F, Paterson AL, Grehan N, Dwerryhouse S, Lao-Sirieix P, Caldas C, Fitzgerald RC. Mechanisms and sequelae of E-cadherin silencing in hereditary diffuse gastric cancer. *J Pathol* 2008;216:295–306.
 - 40 Oliveira C, Sousa S, Pinheiro H, Karam R, Bordeira-Carrico R, Senz J, Kaurah P, Carvalho J, Pereira R, Gusmao L, Wen X, Cipriano MA, Yokota J, Carneiro F, Huntsman D, Seruca R. Quantification of epigenetic and genetic 2nd hits in CDH1 during hereditary diffuse gastric cancer syndrome progression. *Gastroenterology* 2009;136:2137–48.
 - 41 Humar B, Blair V, Charlton A, More H, Martin I, Guilford P. E-cadherin deficiency initiates gastric signet-ring cell carcinoma in mice and man. *Cancer Res* 2009;69:2050–6.
 - 42 Grady WM, Willis J, Guilford PJ, Dunbier AK, Toro TT, Lynch H, Wiesner G, Ferguson K, Eng C, Park JG, Kim SJ, Markowitz S. Methylation of the CDH1 promoter as the second genetic hit in hereditary diffuse gastric cancer. *Nat Genet* 2000;26:16–17.
 - 43 Fitzgerald RC, Caldas C. Clinical implications of E-cadherin associated hereditary diffuse gastric cancer. *Gut* 2004;53:775–8.
 - 44 Suriano G, Oliveira C, Ferreira P, Machado JC, Bordin MC, De Wever O, Bruyneel EA, Moguilevsky N, Grehan N, Porter TR, Richards FM, Hruban RH, Roviello F, Huntsman D, Mareel M, Carneiro F, Caldas C, Seruca R. Identification of CDH1 germline missense mutations associated with functional inactivation of the E-cadherin protein in young gastric cancer probands. *Hum Mol Genet* 2003;12:575–82.
 - 45 Suriano G, Seixas S, Rocha J, Seruca R. A model to infer the pathogenic significance of CDH1 germline missense variants. *J Mol Med* 2006;84:1023–31.
 - 46 Figueiredo J, Soderberg O, Simoes-Correia J, Grannas K, Suriano G, Seruca R. The importance of E-cadherin binding partners to evaluate the pathogenicity of E-cadherin missense mutations associated to HDGC. *Eur J Hum Genet* 2013;21:301–9.
 - 47 Simoes-Correia J, Figueiredo J, Lopes R, Stricher F, Oliveira C, Serrano L, Seruca R. E-cadherin destabilization accounts for the pathogenicity of missense mutations in hereditary diffuse gastric cancer. *PLoS ONE* 2012;7:e33783.
 - 48 Sanchez JM, Figueiredo J, Fonseca M, Duraes C, Melo S, Esmenio S, Seruca R. Quantification of mutant E-cadherin using bioimaging analysis of in situ fluorescence microscopy. A new approach to CDH1 missense variants. *Eur J Hum Genet* 2014. doi:10.1038/ejhg.2014.240
 - 49 Johnston JJ, Rubinstein WS, Facio FM, Ng D, Singh LN, Teer JK, Mullikin JC, Biesecker LG. Secondary variants in individuals undergoing exome sequencing: screening of 572 individuals identifies high-penetrance mutations in cancer-susceptibility genes. *Am J Hum Genet* 2012;91:97–108.
 - 50 Kurian AW, Hare EE, Mills MA, Kingham KE, McPherson L, Whittemore AS, McGuire V, Ladabaum U, Kobayashi Y, Lincoln SE, Cargill M, Ford JM. Clinical evaluation of a multiple-gene sequencing panel for hereditary cancer risk assessment. *J Clin Oncol* 2014;32:2001–9.
 - 51 Eijzena W, Hahn DE, Aaronson NK, Kluijlt I, Bleiker EM. Specific psychosocial issues of individuals undergoing genetic counseling for cancer—a literature review. *J Genet Couns* 2014;23:133–46.
 - 52 Kaurah P, Fitzgerald R, Dwerryhouse S, Huntsman DG. Pregnancy after prophylactic total gastrectomy. *Fam Cancer* 2010;9:331–4.
 - 53 Brandt AC, Tschirgi ML, Ready KJ, Sun C, Darilek S, Hecht J, Arun BK, Lu KH. Knowledge, attitudes, and clinical experience of physicians regarding preimplantation genetic diagnosis for hereditary cancer predisposition syndromes. *Fam Cancer* 2010;9:479–87.
 - 54 Quinn GP, Pal T, Murphy D, Vadaparampil ST, Kumar A. High-risk consumers' perceptions of preimplantation genetic diagnosis for hereditary cancers: a systematic review and meta-analysis. *Genet Med* 2012;14:191–200.
 - 55 Simpson JL. Preimplantation genetic diagnosis at 20 years. *Prenat Diagn* 2010;30:682–95.
 - 56 Gaston D, Hansford S, Oliveira C, Nightingale M, Pinheiro H, Macgillivray C, Kaurah P, Rideout AL, Steele P, Soares G, Huang WY, Whitehouse S, Blowers S, LeBlanc MA, Jiang H, Greer W, Samuels ME, Orr A, Fernandez CV, Majewski J, Ludman M, Dyack S, Penney LS, McMaster CR, Huntsman D, Bedard K. Germline mutations in MAP3K6 are associated with familial gastric cancer. *PLoS Genet* 2014;10:e1004669.
 - 57 Lim YC, di Pietro M, O'Donovan M, Richardson S, Debiram I, Dwerryhouse S, Hardwick RH, Tischkowitz M, Caldas C, Ragnauth K, Fitzgerald RC. Prospective cohort study assessing outcomes of patients from families fulfilling criteria for hereditary diffuse gastric cancer undergoing endoscopic surveillance. *Gastrointest Endosc* 2014;80:78–87.
 - 58 Blair V, Martin I, Shaw D, Winship I, Kerr D, Arnold J, Harawira P, McLeod M, Parry S, Charlton A, Findlay M, Cox B, Humar B, More H, Guilford P. Hereditary diffuse gastric cancer: diagnosis and management. *Clin Gastroenterol Hepatol* 2006;4:262–75.
 - 59 Charlton A, Blair V, Shaw D, Parry S, Guilford P, Martin IG. Hereditary diffuse gastric cancer: predominance of multiple foci of signet ring cell carcinoma in distal stomach and transitional zone. *Gut* 2004;53:814–20.
 - 60 Chun YS, Lindor NM, Smyrk TC, Petersen BT, Burgart LJ, Guilford PJ, Donohue JH. Germline E-cadherin gene mutations: is prophylactic total gastrectomy indicated? *Cancer* 2001;92:181–7.
 - 61 Huntsman DG, Carneiro F, Lewis FR, MacLeod PM, Hayashi A, Monaghan KG, Maung R, Seruca R, Jackson CE, Caldas C. Early gastric cancer in young, asymptomatic carriers of germ-line E-cadherin mutations. *N Engl J Med* 2001;344:1904–9.
 - 62 Lewis FR, Mellinger JD, Hayashi A, Lorelli D, Monaghan KG, Carneiro F, Huntsman DG, Jackson CE, Caldas C. Prophylactic total gastrectomy for familial gastric cancer. *Surgery* 2001;130:612–17; discussion 7–9.
 - 63 Carneiro F, Huntsman DG, Smyrk TC, Owen DA, Seruca R, Pharoah P, Caldas C, Sobrinho-Simoes M. Model of the early development of diffuse gastric cancer in E-cadherin mutation carriers and its implications for patient screening. *J Pathol* 2004;203:681–7.
 - 64 Rogers WM, Dobo E, Norton JA, Van Dam J, Jeffrey RB, Huntsman DG, Kingham K, Chun N, Ford JM, Longacre TA. Risk-reducing total gastrectomy for germline mutations in E-cadherin (CDH1): pathologic findings with clinical implications. *Am J Surg Pathol* 2008;32:799–809.
 - 65 Fujita H, Lennerz JK, Chung DC, Patel D, Deshpande V, Yoon SS, Lauwers GY. Endoscopic surveillance of patients with hereditary diffuse gastric cancer: biopsy recommendations after topographic distribution of cancer foci in a series of 10 CDH1-mutated gastrectomies. *Am J Surg Pathol* 2012;36:1709–17.
 - 66 Shaw D, Blair V, Framp A, Harawira P, McLeod M, Guilford P, Parry S, Charlton A, Martin I. Chromoendoscopic surveillance in hereditary diffuse gastric cancer: an alternative to prophylactic gastrectomy? *Gut* 2005;54:461–8.
 - 67 Richards FM, McKee SA, Rajpar MH, Cole TR, Evans DG, Jankowski JA, McKeown C, Sanders DS, Maher ER. Germline E-cadherin gene (CDH1) mutations predispose to familial gastric cancer and colorectal cancer. *Hum Mol Genet* 1999;8:607–10.
 - 68 Brooks-Wilson AR, Kaurah P, Suriano G, Leach S, Senz J, Grehan N, Butterfield YS, Jeyes J, Schinas J, Bacani J, Kelsey M, Ferreira P, Macgillivray B, MacLeod P, Micek M, Ford J, Foulkes W, Australia K, Greenberg C, LaPointe M, Gilpin C, Nikkel S, Gilchrist D, Hughes R, Jackson CE, Monaghan KG, Oliveira MJ, Seruca R, Gallinger S, Caldas C, Huntsman D. Germline E-cadherin mutations in hereditary diffuse gastric cancer: assessment of 42 new families and review of genetic screening criteria. *J Med Genet* 2004;41:508–17.
 - 69 Hamilton LE, Jones K, Church N, Medlicott S. Synchronous appendiceal and intramucosal gastric signet ring cell carcinomas in an individual with CDH1-associated hereditary diffuse gastric carcinoma: a case report of a novel association and review of the literature. *BMC Gastroenterol* 2013;13:114.

Review

- 70 Oliveira C, Bordin MC, Grehan N, Huntsman D, Suriano G, Machado JC, Kiviluoto T, Aaltonen L, Jackson CE, Seruca R, Caldas C. Screening E-cadherin in gastric cancer families reveals germline mutations only in hereditary diffuse gastric cancer kindred. *Hum Mutat* 2002;19:510–17.
- 71 Molleran V, Mahoney MC. *Breast MRI*. Philadelphia, PA, USA: Elsevier/Saunders, 2014.
- 72 Dunbar K, Canto M. Confocal endomicroscopy. *Curr Opin Gastroenterol* 2008;24:631–7.
- 73 Norton JA, Ham CM, Van Dam J, Jeffrey RB, Longacre TA, Huntsman DG, Chun N, Kurian AW, Ford JM. CDH1 truncating mutations in the E-cadherin gene: an indication for total gastrectomy to treat hereditary diffuse gastric cancer. *Ann Surg* 2007;245:873–9.
- 74 Koea JB, Karpeh MS, Brennan MF. Gastric cancer in young patients: demographic, clinicopathological, and prognostic factors in 92 patients. *Ann Surg Oncol* 2000;7:346–51.
- 75 Stiekema J, Cats A, Kuijpers A, van Coevorden F, Boot H, Jansen EP, Verheij M, Balague Ponz O, Hauptmann M, van Sandick JW. Surgical treatment results of intestinal and diffuse type gastric cancer. Implications for a differentiated therapeutic approach? *Eur J Surg Oncol* 2013;39:686–93.
- 76 Fujioka K. Follow-up of nutritional and metabolic problems after bariatric surgery. *Diabetes Care* 2005;28:481–4.
- 77 Francis WP, Rodrigues DM, Perez NE, Lonardo F, Weaver D, Webber JD. Prophylactic laparoscopic-assisted total gastrectomy for hereditary diffuse gastric cancer. *JSLs* 2007;11:142–7.
- 78 Gotoda T, Yanagisawa A, Sasako M, Ono H, Nakanishi Y, Shimoda T, Kato Y. Incidence of lymph node metastasis from early gastric cancer: estimation with a large number of cases at two large centers. *Gastric Cancer* 2000;3:219–25.
- 79 Choi J, Kim SG, Im JP, Kang SJ, Lee HJ, Yang HK, Kim JS, Kim WH, Jung HC, Song IS. Lymph node metastasis in multiple synchronous early gastric cancer. *Gastrointest Endosc* 2011;74:276–84.
- 80 Kang HJ, Kim DH, Jeon TY, Lee SH, Shin N, Chae SH, Kim GH, Song GA, Kim DH, Srivastava A, Park do Y, Lauwers GY. Lymph node metastasis from intestinal-type early gastric cancer: experience in a single institution and reassessment of the extended criteria for endoscopic submucosal dissection. *Gastrointest Endosc* 2010;72:508–15.
- 81 Roviello F, Rossi S, Marrelli D, Pedrazzani C, Corso G, Vindigni C, Morgagni P, Saragoni L, de Manzoni G, Tomezzoli A. Number of lymph node metastases and its prognostic significance in early gastric cancer: a multicenter Italian study. *J Surg Oncol* 2006;94:275–80; discussion 4.
- 82 Wang G, Ceng G, Zhou B, Chen Y, Jin C, Feng X. Meta-analysis of two types of digestive tract reconstruction modes after total gastrectomy. *Hepatogastroenterology* 2013;60:1817–21.
- 83 Hackenson D, Edelman DA, McGuire T, Weaver DW, Webber JD. Prophylactic laparoscopic gastrectomy for hereditary diffuse gastric cancer: a case series in a single family. *JSLs* 2010;14:348–52.
- 84 Carneiro F, Charlton A, Huntsman D. Hereditary diffuse gastric cancer. In: Bosman FT, ed. *WHO classification of tumours of the digestive system*. 4th edn. IARC Sci Publ, 2010:59–63.
- 85 Dhinra S, Wang H. Nonneoplastic signet-ring cell change in gastrointestinal and biliary tracts: a pitfall for overdiagnosis. *Ann Diagn Pathol* 2011;15:490–6.
- 86 Thompson IW, Day DW, Wright NA. Subnuclear vacuolated mucous cells: a novel abnormality of simple mucin-secreting cells of non-specialized gastric mucosa and Brunner's glands. *Histopathology* 1987;11:1067–81.
- 87 Rubio CA. Five types of pyloric cells in the antral mucosa of the stomach. *Pathol Res Pract* 1992;188:157–61.
- 88 Wang K, Weinrach D, Lal A, Musunuri S, Ramirez J, Ozer O, Keh P, Rao MS. Signet-ring cell change versus signet-ring cell carcinoma: a comparative analysis. *Am J Surg Pathol* 2003;27:1429–33.
- 89 Bosman FT, Carneiro F, Hruban RH, Theise ND. *WHO classification of tumours of the digestive system*. 4th edn. IARC Sci Publ, 2010.
- 90 Lauren P. The two histological main types of gastric carcinoma: diffuse and so-called intestinal-type carcinoma. An attempt at a histo-clinical classification. *Acta Pathol Microbiol Scand* 1965;64:31–49.
- 91 Pandalai PK, Lauwers GY, Chung DC, Patel D, Yoon SS. Prophylactic total gastrectomy for individuals with germline CDH1 mutation. *Surgery* 2011;149:347–55.
- 92 Bardram L, Hansen TV, Gerdes AM, Timshel S, Friis-Hansen L, Federspiel B. Prophylactic total gastrectomy in hereditary diffuse gastric cancer: identification of two novel CDH1 gene mutations—a clinical observational study. *Fam Cancer* 2014;13:231–42.
- 93 Carneiro F, Oliveira C, Suriano G, Seruca R. Molecular pathology of familial gastric cancer, with an emphasis on hereditary diffuse gastric cancer. *J Clin Pathol* 2008;61:25–30.
- 94 Mills SE. *Histology for pathologists*. 3rd edn. Philadelphia, USA: Lippincott Williams & Wilkins, 2007.
- 95 Keller G, Vogelsang H, Becker I, Hutter J, Ott K, Candidus S, Grundei T, Becker KF, Mueller J, Siewert JR, Hofler H. Diffuse type gastric and lobular breast carcinoma in a familial gastric cancer patient with an E-cadherin germline mutation. *Am J Pathol* 1999;155:337–42.
- 96 Lee AF, Rees H, Owen DA, Huntsman DG. Periodic acid-schiff is superior to hematoxylin and eosin for screening prophylactic gastrectomies from CDH1 mutation carriers. *Am J Surg Pathol* 2010;34:1007–13.
- 97 Humar B, Fukuzawa R, Blair V, Dunbier A, More H, Charlton A, Yang HK, Kim WH, Reeve AE, Martin I, Guilford P. Destabilized adhesion in the gastric proliferative zone and c-Src kinase activation mark the development of early diffuse gastric cancer. *Cancer Res* 2007;67:2480–9.
- 98 Mahmud N, Ford JM, Longacre TA, Parent R, Norton JA. Metastatic lobular breast carcinoma mimicking primary signet ring adenocarcinoma in a patient with a suspected CDH1 mutation. *J Clin Oncol* 2015;33:e19–21.
- 99 Koyama T, Sekine S, Taniguchi H, Tsuda H, Ikegami M, Hano H, Kushima R. Hepatocyte nuclear factor 4A expression discriminates gastric involvement by metastatic breast carcinomas from primary gastric adenocarcinomas. *Hum Pathol* 2011;42:1777–84.
- 100 van der Post RS, Bult P, Vogelaa IP, Ligtenberg MJ, Hoogerbrugge N, van Krieken JH. HNF4A immunohistochemistry facilitates distinction between primary and metastatic breast and gastric carcinoma. *Virchows Arch* 2014;464:673–9.
- 101 Worster E, Liu X, Richardson S, Hardwick RH, Dwerryhouse S, Caldas C, Fitzgerald RC. The impact of prophylactic total gastrectomy on health-related quality of life: a prospective cohort study. *Ann Surg* 2014;260:87–93.
- 102 Friess H, Bohm J, Muller MW, Glasbrenner B, Riepl RL, Malfertheiner P, Buchler MW. Maldigestion after total gastrectomy is associated with pancreatic insufficiency. *Am J Gastroenterol* 1996;91:341–7.
- 103 Sakuta H, Suzuki T, Yasuda H, Wakiyama H, Hase K. Plasma vitamin B12, folate and homocysteine levels in gastrectomized men. *Clin Nutr* 2005;24:244–9.
- 104 Ruz M, Carrasco F, Rojas P, Codoceo J, Inostroza J, Basfi-Fer K, Valencia A, Csendes A, Papapietro K, Pizarro F, Olivares M, Westcott JL, Hambidge KM, Krebs NF. Heme- and nonheme-iron absorption and iron status 12 mo after sleeve gastrectomy and Roux-en-Y gastric bypass in morbidly obese women. *Am J Clin Nutr* 2012;96:810–17.
- 105 Adachi S, Kawamoto T, Otsuka M, Todoroki T, Fukao K. Enteral vitamin B12 supplements reverse postgastrectomy B12 deficiency. *Ann Surg* 2000;232:199–201.
- 106 Tondapu P, Provost D, Adams-Huet B, Sims T, Chang C, Sakhaee K. Comparison of the absorption of calcium carbonate and calcium citrate after Roux-en-Y gastric bypass. *Obes Surg* 2009;19:1256–61.
- 107 Mine S, Sano T, Tsutsumi K, Murakami Y, Ehara K, Saka M, Hara K, Fukagawa T, Udagawa H, Katai H. Large-scale investigation into dumping syndrome after gastrectomy for gastric cancer. *J Am Coll Surg* 2010;211:628–36.
- 108 Paik CN, Choi MG, Lim CH, Park JM, Chung WC, Lee KM, Jun KH, Song KY, Jeon HM, Chin HM, Park CH, Chung IS. The role of small intestinal bacterial overgrowth in postgastrectomy patients. *Neurogastroenterol Motil* 2011;23:e191–6.
- 109 Miholic J, Meyer HJ, Muller MJ, Weimann A, Pichlmayr R. Nutritional consequences of total gastrectomy—the relationship between mode of reconstruction, postprandial symptoms, and body-composition. *Surgery* 1990;108:488–94.
- 110 No Stomach For Cancer. <http://www.nostomachforcancer.org/>
- 111 DeGregorio Family Foundation for Stomach and Esophageal Cancer Research. <http://www.degregorio.org/>
- 112 Stichting CDH1. <http://stichting-cdh1.nl/>



Hereditary diffuse gastric cancer: updated clinical guidelines with an emphasis on germline *CDH1* mutation carriers

Rachel S van der Post, Ingrid P Vogelaar, Fátima Carneiro, Parry Guilford, David Huntsman, Nicoline Hoogerbrugge, Carlos Caldas, Karen E Chelcun Schreiber, Richard H Hardwick, Margreet G E M Ausems, Linda Bardram, Patrick R Benusiglio, Tanya M Bisseling, Vanessa Blair, Eveline Bleiker, Alex Boussioutas, Annemieke Cats, Daniel Coit, Lynn DeGregorio, Joana Figueiredo, James M Ford, Esther Heijkoop, Rosella Hermens, Bostjan Humar, Pardeep Kaurah, Gisella Keller, Jennifer Lai, Marjolijn J L Ligtenberg, Maria O'Donovan, Carla Oliveira, Hugo Pinheiro, Krish Ragunath, Esther Rasenberg, Susan Richardson, Franco Roviello, Hans Schackert, Raquel Seruca, Amy Taylor, Anouk ter Huurne, Marc Tischkowitz, Sheena Tjon A Joe, Benjamin van Dijck, Nicole C T van Grieken, Richard van Hillegersberg, Johanna W van Sandick, Rianne Vehof, J Han van Krieken and Rebecca C Fitzgerald

J Med Genet 2015 52: 361-374 originally published online May 15, 2015
doi: 10.1136/jmedgenet-2015-103094

Updated information and services can be found at:
<http://jmg.bmj.com/content/52/6/361>

These include:

Supplementary Material

Supplementary material can be found at:
<http://jmg.bmj.com/content/suppl/2015/05/15/jmedgenet-2015-103094.DC1.html>

References

This article cites 102 articles, 21 of which you can access for free at:
<http://jmg.bmj.com/content/52/6/361#BIBL>

Open Access

This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Email alerting service

Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Topic Collections

Articles on similar topics can be found in the following collections

[Open access](#) (149)
[Breast cancer](#) (230)
[Epidemiology](#) (614)

To request permissions go to:
<http://group.bmj.com/group/rights-licensing/permissions>

To order reprints go to:
<http://journals.bmj.com/cgi/reprintform>

To subscribe to BMJ go to:
<http://group.bmj.com/subscribe/>

Notes

To request permissions go to:
<http://group.bmj.com/group/rights-licensing/permissions>

To order reprints go to:
<http://journals.bmj.com/cgi/reprintform>

To subscribe to BMJ go to:
<http://group.bmj.com/subscribe/>